Effects of Insulin on the Disposal of ¹⁴C-Labelled Very Low Density Lipoprotein Triglycerides in Intact and Hepatectomized Rats

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Summary. In sham-operated rats, intravenous administration of ¹⁴C-very low density lipoprotein triglycerides (with labelled esterified fatty acids) caused an initial decrease and subsequent increase in plasma ¹⁴C-lipids of both very low density lipoproteins (VLDL) (density < 1.006) and lipoproteins of density > 1.019. There was a similar change in ¹⁴C-lipids in adipose tissue and heart whereas in kidney, spleen and liver, ¹⁴C-lipids increased initially and then decreased. Insulin treatment in sham-operated animals decreased circulating ¹⁴C-lipids in VLDL and in lipoproteins of density > 1.019, while intermediate density (1.006–1.019) lipoproteins increased. Insulin also enhanced the radioactivity retained in spleen. In functionally hepatectomized rats, ¹⁴C-lipids progressively increased in heart. Insulin treatment in these rats enhanced the

Insulin administration is known to reduce the elevated levels of circulating triglycerides and very low density lipoprotein (VLDL)-triglycerides in diabetic subjects [1] and it is well established that insulin enhances the deposition of triacyglycerol fatty acids in adipose tissue by increasing its lipoprotein lipase activity [2-4]. There are, however, situations in which insulin excess is associated with augmented circulating triglyceride concentrations [5-7], and the effects of insulin on lipoprotein lipase activity in other extrahepatic tissues do not follow the changes found in the adipose tissue enzyme [8, 9]. As insulin also stimulates the secretion of newly synthesized VLDL-triglycerides by liver preparations [10, 11], changes in circulating lipoproteins 'in vivo' produced by insulin are the result of a combination of hepatic and extrahepatic effects. In the present study, disassociation of these effects was achieved by determining the changes produced by insulin on the disposal of prelabelled VLDL-triglycerides administered to functionally hepatectomized rats and to sham-operated controls, to determine the effects of the hormone 'in vivo' on extrahepatic VLDL-triglyceride catabolism.

disappearance from circulation of ¹⁴C-VLDL and of lipoproteins of density > 1.019, as well as the appearance of ¹⁴C-intermediate density lipoproteins. The appearance of ¹⁴C-lipids in white adipose tissue also was augmented, while it decreased in heart and lung. Thus, in sham-operated animals, insulin apparently stimulates the uptake of products of VLDL metabolism by cells in the reticuloendothelial system, while in functionally hepatectomized rats there is increased heart utilization of VLDL triglycerides, and insulin enhances the net extrahepatic catabolism of these lipoproteins.

Key words: Plasma lipoproteins, rat, insulin, spleen, hepatectomy.

Materials and Methods

Female Wistar rats weighing 160-190 g and fed standard rat chow were maintained in a temperature $(22 \pm 2 \,^{\circ}C)$ and light cycle (light from 9.00 to 21.00 h) controlled room. Animals were subjected to functional hepatectomy under sodium pentobarbital anaesthesia (40 mg/kg, IP) between 10.00 and 12.00 h, following the method of Russell [12], in which blood vessels supplying the liver (the coeliac axis and portal vein just above the first division to the hepatic lobes) were tied. The medium and left lateral hepatic lobes were ligated [13] and the abdominal wall closed with wound clips. The liver was left in situ but proved to be entirely excluded from circulation as < 0.09% of injected isotope was recovered in liver 30 min after injection through the inferior vena cava of 1-14C-palmitate bound to 8% bovine albumin, which is known to be taken up readily by the liver [14]. The entire surgical procedure took < 15 min. In sham-operated control rats studied in parallel, laparotomy and handling of the liver were performed as in experimental animals but without tying any vessel.

Immediately after surgery, animals were injected through the inferior vena cava with 1 IU/kg body weight Actrapid-monocomponent insulin (Novo Industri, Copenhagen, Denmark) or with 0.9% NaCl, followed 5 min later by 0.7 ml of rat ¹⁴C-labelled VLDL-triglycerides containing 0.35 mg of triacyglycerols, prepared as described below. Animals were sacrificed 5, 15, or 30 min thereafter and blood, lumbar fat pads, kidneys, spleen, liver, heart, lungs, and skeletal muscle from one leg immediately collected. Aliquots of plasma samples were used for counting radioactivity and for purification of lipids [15].

Table 1. Plasma ¹⁴C-lipids after the intravenous administration of ¹⁴C-VLDL-triglycerides in the rat treated or not with insulin (1 IU/kg)

	Untreated animals	3		Treated with insulin			
	VLDL (density < 1.006)	IDL (density > 1.006 < 1.019)	LDL+HDL (density >1.019)	VLDL (density < 1.006)	IDL (density > 1.006 < 1.019)	LDL+HDL (density > 1.019)	
Sham-or	perated control rats						
5 min	7153 ± 1347	960 ± 113	4404 ± 1016	7415 ± 2054	1601 ± 499	4455 ± 1018	
15 min	$2963 \pm 363^{\circ}$	1273 ± 192	$1838 \pm 256^{\circ}$	$1264 \pm 261^{2a, c}$	2069 ± 286^{a}	$1096 \pm 291^{\circ}$	
30 min	4045 ± 442	1030 ± 173	3877 ± 940	$1215 \pm 107^{3a, c}$	1066 ± 171	$716 \pm 114^{a, 2c}$	
Hepatec	tomized rats						
5 min	8378 ± 819	1244 ± 150	4638 ± 713	7831 ± 760	1437 ± 269	4516 ± 790	
15 min	6406 ± 696^{2b}	1684 ± 233	4805 ± 662	$4378 \pm 918^{b, c}$	$1843 \pm 217^{\circ}$	$1196 \pm 333^{2a, 2c}$	
30 min	4639 ± 475^{2c}	1208 ± 114	3773 ± 297	$3038 \pm 270^{a, 3b, 3c}$	$1899 \pm 231^{a, b}$	$2284 \pm 208^{a, 3b, c}$	

Results are expressed as disintegrations $\cdot \min^{-1} \cdot ml^{-1}$ (mean ± SEM, n = 5). Significance of difference values are as follows: ^ainsulin treated versus untreated rats; ^bhepatectomized versus sham-operated control rats; ^cversus corresponding 5 min values. ^{a. b. c} = p < 0.05; ^{2a, 2b, 2c} = p < 0.01; ^{3a, 3b, 3c} = p < 0.001

Other plasma aliquots were used for lipoprotein fractionation by sequential flotation in a Beckman L5-75 ultracentrifuge (Beckman Instruments, Palo Alto, California, USA) as described previously [16].

Rat ¹⁴C-labelled VLDL-triglycerides were obtained by IV injection into the tail of other female rats (185–190 g body weight) of 1-14Cpalmitate (100 µCi, 51.8 mCi/mmol, Radiochemical Centre, Amersham, Bucks, UK) as described elsewhere [17]. Animals were sacrificed 20 min later under light ether anaesthesia, and blood was collected from the aorta into a syringe containing 100 mg of Na₂-EDTA. Plasma was centrifuged under 0.15 mmol/l NaCl (d = 1.006) containing 1 mmol/l EDTA for 18 h at 143,000 g and 15 °C in a Beckman L5-75 ultracentrifuge. Supernatants were collected with a tube slicer and were purified further by dialysis against three changes of 0.9% NaCl (5l) for a total of 18 h at 4 °C. This preparation contained particles of diameter (150-600) Å when observed under electronmicroscopy [18] and was characterized as rat ¹⁴C-VLDL-triglycerides, since it contained only pre-beta lipoproteins on agarose gel electrophoresis [16] and more than 91% of the label appeared as esterified fatty acids of neutral lipids (presumably triglycerides) on fractionation [19]. Radioactivity was measured in a PPO/POPOP scintillation cocktail dissolved in xylene and Triton X-100 and using a Nuclear-Chicago counter (Isocap 300) provided with an external standard device. Radioactive measurements were adjusted to 10⁶ d.p.m. for the ¹⁴C-labelled VLDL preparation injected into each animal. Statistical comparison between groups was performed with the Student's t-test.

Results

Values of ¹⁴C-lipids in plasma lipoprotein fractions at different times after the IV administration of ¹⁴C-VLDL-triglycerides are summarized in Table 1. In sham-operated control rats not receiving insulin, radioactivity in lipids of plasma VLDL (d < 1.006) and low density lipoproteins plus high density lipoproteins (LDL + HDL) (d > 1.019) fractions decreased from 5 to 15 min after tracer injection (p < 0.05 for all 15-min versus corresponding 5-min values) but at 30 min, values did not differ significantly from those at 5 min. In contrast, radioactivity in plasma as intermediate density lipoproteins (IDL) (d > 1.006, < 1.019) did not change after tracer injection. Prior insulin treatment did not modify the distribution of label in plasma lipoproteins at 5 min but later it reduced the radioactivity in lipids of both VLDL and LDL + HDL, the difference from untreated animals being statistically significant at 15 and 30 min for VLDL and at 30 min for LDL + HDL (Table 1). On the contrary, insulin treatment enchanced ¹⁴C-IDL values at 15 min versus values at that time in untreated animals (p < 0.05), and this difference disappeared at 30 min (Table 1). When the same experiments were performed in hepatectomized animals not receiving insulin, values of plasma radioactive lipoprotein fractions did not differ from those in sham-operated rats at 5 min. Unlike sham-operated untreated rats, in the hepatectomized animals, there was no change in plasma radioactivity in lipoprotein fractions at 15 min, thus values of VLDL and LDL + HDL were significantly higher than those of sham-operated basal values at this time. These differences disappeared at 30 min due to the rise in values of sham-operated untreated rats and the very small changes in hepatectomized animals. Treatment of hepatectomized rats with insulin showed that radioactivity in lipids in VLDL at 30 min, and in LDL + HDL at 15 and 30 min was significantly reduced, while in IDL it was increased at 30 min compared with values in hepatectomized untreated animals.

At each of the times studied, animals were sacrificed and ¹⁴C-lipids determined in organs (Table 2). Per unit of wet weight, the greatest radioactivity in lipids of organs from sham-operated untreated animals was found at 5 min in liver, followed by spleen, lung, heart, skeletal muscle, white adipose tissue, and kidney. In spleen, liver, and kidney, ¹⁴C-lipids significantly increased from 5 to 15 min, this change being particularly manifest in the spleen (p < 0.001; Table 2). From 15 to 30 min, ¹⁴C-lipids in these three organs decreased, values in liver falling to the 5 min value, while those in spleen and kidney remained significantly augmented. Variations of ¹⁴Clipids in adipose tissue, heart and lung with time were opposite to those in spleen, liver, and kidney, although they were statistically significant for heart only (Table 2).

In skeletal muscle, ¹⁴C-lipids did not change at the times studied. Insulin treatment in these sham-operated rats enhanced the appearance of ¹⁴C-lipids in adipose

Table 2. Tissue ¹⁴C-lipids after the intravenous administration of ¹⁴C-VLDL-triglycerides in sham-operated or hepatectomized rats treated or not with insulin (1 IU/kg)

	Adipose tissue	Kidney	Skeletal muscle	Spleen	Liver	Heart	Lung
Sham opera	ted rats						
Untreated							
5 min	3056 ± 556	2254 ± 438	5200 ± 727	13226 ± 1970	31448 ± 3718	7093 ± 448	7809 ± 1259
15 min	2128 ± 335	5366 ± 412^{3c}	5217 ± 718	40393 ± 4456^{3c}	$47327 \pm 5341^{\circ}$	$3571 \pm 936^{\circ}$	6787 ± 1197
30 min	4307 ± 854	$4256 \pm 568^{\circ}$	4050 ± 662	22921 ± 1503^{2c}	26881 ± 1637	4175 ± 574^{2c}	7009 ± 1232
Insulin trea	ted						
5 min	4878 ± 804	3381 ± 945	4581 ± 605	17236 ± 1785	36336 ± 3351	6139 ± 1273	8177 ± 1540
15 min	4643 ± 504^{2a}	3738 ± 302^{a}	4787 ± 617	42745 ± 1663^{3c}	51478 ± 3038^{2c}	$3093 \pm 231^{\circ}$	6130 ± 410
30 min	5018 ± 961	3699 ± 730	4647 ± 532	$35460 \pm 1621^{3a,3}$	$3^{\circ} 29698 \pm 2345$	3452 ± 696	5343 ± 806
Hepatectom	ized rats						
Untreated							
5 min	3827 ± 709	2506 ± 705	7206 ± 1770	241 ± 122^{3b}	242 ± 76^{3b}	7204 ± 2390	9994 ± 2552
15 min	3202 ± 299^{b}	4947 ± 1118	5215 ± 1502	219 ± 35^{3b}	189 ± 42^{3b}	8552 ± 1622	9869 ± 1642
30 min	2973 ± 433	5585 ± 780^{2c}	5154 ± 1103	347 ± 42^{3b}	517 ± 60^{3b}	$12408 \pm 1433^{3\text{h,c}}$	11118 ± 888^{b}
Insulin trea	ted						
5 min	4052 ± 885	4028 ± 820	6594 ± 1854	390 ± 125^{3b}	280 ± 51^{3b}	6618 ± 1335	8325 ± 1581
15 min	3402 ± 318	4531 ± 603	4175 ± 1108	282 ± 68^{3b}	479 ± 129^{3b}	$7775\pm1739^{\mathrm{b}}$	7085 ± 609
30 min	5222 ± 318^{2a}	4521 ± 603	4226 ± 974	232 ± 59^{3b}	335 ± 60^{3b}	$8490 \pm 676^{3b, a}$	6396 ± 1696^{a}

Results are expressed as disintegrations $\cdot \min^{-1} \cdot g$ wet weight⁻¹ (mean ± SEM, n = 5). Significance of difference values are as follows: ^ainsulintreated versus untreated rats; ^bhepatectomized versus sham-operated control rats; ^cversus corresponding 5 min values. ^{a, b, c} = p < 0.05; ^{2a, 2b, 2c} = p < 0.01; ^{3a, 3b, 3c} = p < 0.001

tissue and reduced it in kidney at 15 min, while a marked effect was produced in spleen, where the disappearance of ¹⁴C-lipids occurring between 15 and 30 min in untreated animals decreased (Table 2). In hepatectomized animals, ¹⁴C-lipid values in adipose tissue, kidney and skeletal muscle were similar to those in shamoperated animals (Table 2). On the contrary, radioactivity in both spleen and liver lipids was always < 0.06%/gof the administered tracer, indicating the efficiency of hepatectomy (Table 2). Although ¹⁴C-lipid values in heart and lung were the same in hepatectomized and sham-operated animals at 5 min, they rose subsequently in the former versus the latter group, the difference being statistically significant at 15 min for heart and at 30 min for both heart and lung (Table 2). At 30 min, insulin treatment significantly increased ¹⁴C-lipids in white adipose tissue while decreasing them in heart and lung (Table 2).

Discussion

The appearance in plasma of radioactive IDL and lipoproteins of d > 1.019 (LDL + HDL) 5 min after intravenous administration of ¹⁴C-VLDL-triglycerides in sham-operated rats corresponds to the known short (2–5 min) half-life of VLDL [20, 21]. The subsequent disappearance from circulation of both ¹⁴C-VLDL and ¹⁴C-(LDL + HDL) coincided with a great increase in radioactivity in spleen and liver, and these results may be due to reticuloendothelial tissue uptake of products of extrahepatic VLDL metabolism such as VLDL rem-

nants and LDL. It has been shown recently that VLDL remnants are efficiently taken up by liver [21] and that macrophages and reticuloendothelial tissues contribute substantially to receptor-independent LDL catabolism [22, 23]. Retention of radioactivity by the spleen after insulin treatment coincided with a decrease in circulating ¹⁴C-(LDL + HDL) demonstrating for the first time that insulin stimulates the uptake of VLDL metabolism products (presumably LDL) by cells of the reticuloendothelial system. This finding may be important in view of present evidence that hyperinsulinaemia is a risk factor for atherosclerosis [24].

In functionally hepatectomized rats, the appearance of ¹⁴C-lipids in plasma IDL and LDL + HDL after administration of ¹⁴C-VLDL-triglycerides demonstrates the capacity of the rat to degrade VLDL to lipoproteins of higher density without the intervention of the liver, in agreement with Suri et al. [25]. The conversion of VLDL to IDL is catalyzed by the action of extrahepatic lipoprotein lipase [26], releasing non-esterified fatty acid (NEFA) from the hydrolysis of VLDL-triglycerides which are taken up by the subjacent tissue. After functional hepatectomy in the rat, this process seems to be enhanced in heart, and less markedly so in lung, as shown by the rise in ¹⁴C-lipids in these tissues after administration of ¹⁴C-VLDL-triglycerides. Increased formation of glyceride glycerol from administered glycerol has been reported previously in the heart of hepatectomized-nephrectomized rats [27]. All these data indicate that hepatectomy enhances heart and lung lipoprotein lipase activities and that NEFA released from VLDL-triglyceride hydrolysis are subsequently taken up by these tissues and re-esterified with α -glycerol phosphate formed from circulating glycerol or other substrates (i.e., glucose). This interpretation is in agreement with the observed insulin effect decreasing ¹⁴Clipid appearance in these tissues in hepatectomized rats. since this hormone is known to inhibit lipoprotein lipase activity in heart [8, 9] and has been shown to decrease the incorporation of circulating glycerol into lipids of both heart and lung in hepatectomized rats [28]. These negative effects of insulin on the uptake of fatty acids from circulating VLDL-triglycerides in heart and lung were overcompensated for by opposite effects in other extrahepatic tissues, as shown by the accelerated disappearance of ¹⁴C-VLDL-triglycerides from the circulation and the enhanced levels ¹⁴C-IDL in functionally hepatectomized rats. The insulin effect of increasing the appearance of ¹⁴C-lipids in adipose tissue in these animals is in agreement with the hormonal effect of lipoprotein lipase enhancement in this tissue [2-4, 8, 29]. The latter, together with the relatively large mass of adipose tissue indicate that it is the main tissue responsible for the enhanced disappearance from circulation of VLDL-triglycerides produced by insulin in functionally hepatectomized rats.

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References

- 1. Nikkilä EA, Hormila P (1978) Serum lipids and lipoproteins in insulin-treated diabetes. Demonstration of increased high density lipoprotein concentrations. Diabetes 27: 1078–1086
- Garfinkel AS, Nilsson-Ehle P, Schotz MC (1976) Regulation of lipoprotein lipase induction by insulin. Biochim Biophys Acta 424: 264–273
- Desai R, Hollenberg CH (1975) Regulation by insulin of lipoprotein lipase and phosphodiesterase activities in rat adipose tissue. Isr J Med Sci 11: 540–550
- Parkin SM, Walker K, Ashby P, Robinson DS (1980) Effect of glucose and insulin on the activation of lipoprotein lipase and on protein synthesis in rat adipose tissue. Biochem J 188: 193–199
- Eaton P, Nye WHR (1975) The relationship between insulin secretion and triglyceride concentrations in endogenous lipemia. J Lab Clin Med 81: 682–695
- Tobey TA, Greenfield M, Kraemer F, Reaven GM (1980) Relationship between insulin resistance, insulin secretion, very low density lipoprotein, and plasma triglyceride levels in normotriglyceridemic man. Metabolism 30: 165–171
- Chait A, Janus E, Stuart Mason A, Lewis B (1979) Lipodystrophy with hyperlipidaemia: the role of insulin in very low density lipoprotein over-synthesis. Clin Endocrinol 10: 173–178
- Borensztajn J, Samols DR, Rubenstein AH (1972) Effects of insulin on lipoprotein lipase activity in the rat heart and adipose tissue. Am J Physiol 223: 1271–1275
- Friedman G, Stein O, Stein Y (1978) Lipoprotein lipase of cultured mesenchymal rat heart cells. III. Effect of glucocorticoids and insulin on enzyme formation. Biochim Biophys Acta 531: 222-232
- 10. Topping DL, Mayes PA (1972) The immediate effects of insulin and fructose on the metabolism of the perfused liver. Changes in

lipoprotein secretion, fatty acid oxidation and esterification, lipogenesis and carbohydrate metabolism. Biochem J 126: 295-311

- 11. Beynen AC, Haagsman HP, Van Golde LMG, Geelen MJH (1981) The effects of insulin and glucagon on the release of triacylglycerols by isolated rat hepatocytes are mere reflections of the hormonal effects on the rate of triacylglycerol synthesis. Biochim Biophys Acta 665: 1–7
- 12. Russell JA (1942) The anterior pituitary in the carbohydrate metabolism of the eviscerated rat. Am J Physiol 136: 95-104
- Higgins GM, Anderson RM (1931) Experimental pathology of the liver. I. Restoration of the liver of the white rat following partial surgical removal. Arch Pathol 12: 186–195
- Carmaniu S, Herrera E (1979) Effect of evisceration on the disposal of ¹⁴C-palmitate in the rat. Arch Intern Physiol Biochim 87: 955–961
- Folch JM, Lee M, Sloane-Stanley GH (1957) A simple method for the isolation and purification of total lipids from animal tissues. J Biol Chem 226: 497–509
- 16. Argilés J, Herrera E (1981) Lipids and lipoproteins in maternal and fetus plasma in the rat. Biol Neonate 39: 37-44
- Carmaniu S, Herrera E (1979) Conversion of (U-¹⁴C)-glycerol, (2-³H)-glycerol and (1-¹⁴C)-palmitate into circulating lipoproteins in the rat. Rev Esp Fisiol 35: 459–464
- Lasunción MA, Llobera M, Herrera E (1981) Morphological and compositional changes of rat plasma triglyceride-rich lipoproteins incubated with adipose tissue. Arch Int Physiol Biochim 89: 57-62
- Kerpel S, Shafrir E, Shapiro B (1961) Mechanism of fatty acid assimilation in adipose tissue. Biochem Biophys Acta 46: 495–504
- Faergeman O, Havel RJ (1975) Metabolism of cholesteryl esters of rat very low density lipoproteins. J Clin Invest 55: 1210–1218
- Windler E, Chao Y, Havel RJ (1980) Determinants of hepatic uptake of triglyceride-rich lipoproteins and their remnants in the rat. J Biol Chem 255: 5475-5480
- Slater HR, Packard CJ, Shepherd J (1982) Receptor independent catabolism of low density lipoprotein. Involvement of the reticuloendothelial system. J Biol Chem 257: 307–310
- 23. Knight B, Soutar AK (1982) Degradation by cultured fibroblasts and macrophages of unmodified and 1,2-cyclohexanedione-modified low-density lipoprotein from normal and homozygous familial hypercholesterolaemic subjects. Biochem J 202: 145–152
- 24. Steiner G (1981) Diabetes and atherosclerosis. An overview. Diabetes 30 (Suppl 2): 1–7
- 25. Suri BS, Targ ME, Robinson DS (1979) The metabolic conversion of very-low-density lipoprotein into low-density lipoprotein by the extrahepatic tissue of the rat. Biochem J 178: 455–466
- Nilsson-Ehle P, Garfinkel AS, Schotz MC (1980) Lipolytic enzymes and plasma lipoprotein metabolism. Ann Rev Biochem 49: 667–693
- Carmaniu S, Herrera E (1980) Extrahepatic utilization of ¹⁴C-glucose and ¹⁴C-glycerol in the eviscerated rat. Diabetes Metab 6: 239–244
- Mampel T, Palacin M, Herrera E (1981) Contraposed effects of insulin on extrahepatic glycerol utilization. Acta Endocrinol 97 (Suppl 243) (Abstract)
- Ashby P, Robinson DS (1980) Effects of insulin, glucocorticoids and adrenaline on the activities of rat adipose-tissue lipoprotein lipase. Biochem J 188: 185–192

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