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***Campylobacter* prevalence and risk factors in poultry slaughterhouses in Spain following the EU criteria**

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10 **ABSTRACT**

11 Background: *Campylobacter* is the main pathogen involved in zoonotic gastrointestinal  
12 diseases. Last year, the European regulation 2017/1495 on *Campylobacter* in broiler  
13 carcasses came into force. In this context, the aim of this study was to assess the potential  
14 risk factors associated with exceeding the 1,000 CFU/g criterion set by the European  
15 Commission in several slaughterhouses in Spain.

16 Methods: Information relating to 12 factors were collected using questionnaires. Samples  
17 were collected from 12 Spanish abattoirs during June, July and August 2017 (n=1,725)  
18 and were analysed following ISO/TS 10272-2:2006 method.

19 Results: The proportion of *Campylobacter*-positive samples was 23.7% (n=409).  
20 Analysis of the flock age (41-50 days) revealed a significantly increased OR in  
21 *Campylobacter* enumeration (OR=7.41). Moreover, scalding temperature (51.9.54 °C)  
22 was positively associated with an increase in OR (OR=2.75). Time in transit to slaughter  
23 (1-1.5h), showed a significant OR decrease (OR=0.25). However, when processed for  
24 more than 2 hours, presented an increase in OR (OR=4.44). Regarding carcass weight,  
25 the range from 3.21-3.58 presented a decrease in OR (OR=0.01).

26 Conclusion: The outcomes of this study suggest that although most chickens are  
27 contaminated by the bacterium, the prevalence that exceeds the limit of 1,000 CFU/ is not  
28 so high as we thought.

29 **Key words:** *Campylobacter*, poultry, slaughterhouse, quantitative, risk factor

## 30 INTRODUCTION

31 *Campylobacter* is the main bacterial cause of human gastroenteritis in most industrialised  
32 countries and has been since 2005 [1-3]. In 2017, campylobacteriosis was the most  
33 commonly reported zoonosis, representing almost 70% of all cases [4]. The European  
34 Surveillance System described 246,158 confirmed cases of human campylobacteriosis in  
35 EU. However, their severity in terms of reported case fatality was low (0.04%) [4].

36 Consuming undercooked poultry meat and cross-contamination due to a lack of hygiene  
37 conditions are the main sources of campylobacteriosis outbreaks. Several studies have  
38 shown *Campylobacter* prevalence in poultry carcasses. Lawes et al. (2012) observed that  
39 70% of poultry carcasses in the UK were contaminated with the bacterium [5]. Moreover,  
40 a qualitative cross-contamination study showed that *Campylobacter* was easily  
41 transferred from raw chicken products to cutting boards, plates and particularly to hands  
42 [6].

43 To lower the risk of human infection, in the drive to reduce the number of flocks colonised  
44 with this bacterium most studies have focused on rearing at poultry farm level [7-9].  
45 Nevertheless, the epidemiology of *Campylobacter* in poultry production is still not fully  
46 understood, making it difficult to control [10].

47 In this context, biosecurity plans, immunity and nutritional strategies have been studied.  
48 However, none of these measures has managed to reduce *Campylobacter* prevalence to  
49 acceptable levels [11].

50 Even though *Campylobacter* is the main pathogen involved in zoonotic gastrointestinal  
51 diseases, there is no national or European control programme at farm level, in contrast to  
52 *Salmonella*, probably due to the incomplete knowledge of *Campylobacter* epidemiology.  
53 In this regard, legislators have been working to limit *Campylobacter* presence in broiler  
54 carcasses. To this end, earlier this year, European regulation 2017/1495 on

55 *Campylobacter* in broiler carcasses came into force. Under this new European regulation,  
56 neck skin from broiler carcasses at the slaughterhouse must be analysed for  
57 *Campylobacter* after chilling with the microbiological criterion of maximum 1,000  
58 CFU/g (colony form units/per gram) [12].

59 It has been suggested that the pre-slaughter handling and transport of broiler chickens are  
60 important stress factors for bacteria shedding [13]. Poultry carcasses could be  
61 contaminated with the bacterium due to splashing of intestinal contents during the  
62 slaughtering process. Mainly the stages of scalding, de-feathering and evisceration could  
63 increase the contamination of carcasses [14,15], representing critical points where the  
64 *Campylobacter* counts can be reduced.

65 In this context, the aim of this study was to assess the potential risk factors associated  
66 with exceeding the 1,000 CFU/g criterion set by the European Commission in several  
67 slaughterhouses in Spain.

68 **MATERIAL AND METHODS**

69 *Study sample*

70 This study was carried out on 12 Spanish commercial broiler poultry processing plants  
71 which slaughter close to 20% of the chickens reared in Spain. The processing line was  
72 operating under standard commercial conditions. Over three months (from June to August  
73 2017), three visits were made to each processing plant, with a total of 36 visits. At each  
74 visit, the first and the last flock processed were sampled in order to assess cross-  
75 contamination. Characteristics of the slaughterhouses are listed in Table 1.

76 **Table 1.** Characteristics of each slaughterhouse.

Variable	Slaughterhouse											
	1	2	3	4	5	6	7	8	9	10	11	12
<b>Number of animals slaughtered (total)</b>	10,001-30,000	1,000-30,000	1,000-10,000	1,000-10,000	20,001-30,000	1,000-10,000	30,000-40,000	80,001-100,000	20,001-30,000	10,001-30,000	80,001-100,000	1000-10,000
<b>Flock size animals</b>	4,001-5,000	4,001-5,000	500-2,500	1,001-2,000	2501-6500	500-6,500	2501-6500	4,501-8550	4501-8550	500-8500	2501-6500	500-2,500
<b>Slaughter age</b>	41-50	41-50	71-80	71-100	41-50	41-60	30-50	30-50	30-60	30-50	30-60	81-90
<b>Scalding temperature (°C)</b>	48-51.8	48-51.8	48-51.8	48-51.8	48-51.8	48-54	48-54	51.9-54	48-54	48-54	51.8-54	48-51.8
<b>Amount of chlorine (ppm)</b>	0-0.5	0-0.5	0.5	0.5	0.51-0.95	0.51-0.95	0.51-0.95	0.51-0.95	0-0.95	0-0.5	0-0.5	0-0.5
<b>Conservation temperature (°C)</b>	≤0	≤0	≤0	≤0	>0	>0	>0	≤0	>0	>0	>0	≤0
<b>Chilling time (min)</b>	60-100	151-200	>300	251->300	60-100	60-300	101-150	101-150	101-150	60-100	201-250	60-100
<b>Time in transit to slaughter (h)</b>	1.6-2	1-1.5	1.6->2	1-1.5	<1	1-2	1-1.5	<1-1.5	<1-2	<1->2	<1	1-1.5
<b>Carcass weight (kg)</b>	1-3	2.51-3	>3	>3	2.01-3	2.01-3	1-3	1-3	2.01->3	1-2.5	2.51->3	>3

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78

### 79 ***Sampling during slaughtering***

80 From each studied flock, samples were collected at five selected stages of the processing  
81 line (after scalding, after de-feathering, after eviscerating, after washing and after  
82 chilling) to determine the degree of *Campylobacter* contamination in the carcasses  
83 throughout the processing line (figure 1). In each stage, five carcasses were collected (25  
84 carcasses per flock), which means 50 carcasses per sampling session, 25 from the first  
85 flock and 25 from the last flock. All samples were transported under cold storage  
86 conditions and analysed within 24h.

87

### 88 ***Carcass sampling***

89 Sampling of carcasses was carried out by collecting neck skin with aseptic conditions and  
90 placing them in an individual sterile bag (Seward, Worthing, UK). Neck skin samples  
91 were obtained from each carcass by removing a strip of neck skin (25g) with a sterile  
92 scalpel and tweezers. Samples were transported to the laboratory on the same day of  
93 sampling. Each individual skin sample was placed in a sterile bag and diluted at 1:10  
94 vol/vol Buffered Peptone Water (BPW, Scharlau®, Barcelona, Spain). The mix was  
95 homogenised by stomaching at 230 rpm for 120 seconds (Stomacher®400 circulator,  
96 Seward Ltd., Worthing, UK). Then, 1 ml of the homogenate was used for *Campylobacter*  
97 enumeration.

98

### 99 ***Campylobacter enumeration***

100 Neck skin homogenates were analysed according to the ISO/TS 10272-2:2017, the  
101 horizontal method for the enumeration of *Campylobacter* in food and feed stuffs [16]. For  
102 detection purposes, serial dilutions were prepared in BPW and a 0.1 mL drop from each  
103 inoculum was plated onto mCCDA (Modified Charcoal Cefoperazone Deoxycholate

104 Agar, Oxoid, Dardilly, France). The samples were then incubated at  $41.5 \pm 1^\circ\text{C}$  in a  
105 microaerobic atmosphere (84%  $\text{N}_2$ , 10%  $\text{CO}_2$ , 6%  $\text{O}_2$ ) for  $44 \pm 4$  hours. Five  
106 *Campylobacter*-like colonies were plated in Columbia blood agar (AES Laboratories®,  
107 Bruz Cedex, France) for further characterisation. Colony morphology and motility were  
108 evaluated under dark field microscopy. Confirmation of the suspicious colonies was  
109 performed by oxidase and catalase tests and plating at different temperatures and  
110 atmospheres ( $41.5^\circ\text{C}$  under microaerophilic conditions and  $25^\circ\text{C}$  under aerobic  
111 conditions) onto Columbia blood agar (AES Laboratories®, Bruz Cedex, France).

112

### 113 ***Study critical points***

114 Every sampling was accompanied by a questionnaire that included the information on the  
115 slaughterhouse, month of sacrifice, position of the batch in the daily slaughter schedule,  
116 processing stage at sampling, total number of animals slaughtered, flock size, age of  
117 animals at slaughter, scalding temperature, amount of chlorine, chilling time and  
118 conservation temperature, time in transit to slaughter and carcass weight. The variable  
119 levels studied are represented in Table 2.



120 **Table 2.** Number of variables assessed during the sampling period.

Variable	Level	Reference
<b>Slaughterhouse</b>	1	Lawes et al. (2012)
	2	
	3	
	4	
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	8	
	9	
	10	
	11	
	12	
<b>Month</b>	June	Lawes et al. (2012)
	July	
	August	
<b>Batch position</b>	1 <sup>st</sup>	Hue et al. (2010)
	2 <sup>nd</sup>	
<b>Processing stages</b>	After scalding	Guerin et al. (2010)
	After de-feathering	
	After eviscerating	
	After washing	
	After chilling	
<b>Number of animals slaughtered (total)</b>	1,000-10,000	
	10,001-20,000	
	20,001-30,000	
	30,001-40,000	
	40,001-80,000	
	80,001-100,000	
<b>Flock Size (animals)</b>	500-2,500	Näther et al. (2009)
	2,501-4,500	
	4,501-6,500	
	6,501-8,550	
<b>Slaughter age (days)</b>	30-40	Näther et al. (2009)
	41-50	
	51-100	
<b>Scalding temperature (°C)</b>	48-51.8	Hue et al. (2010)
	51.9-54	
<b>Amount of chlorine (ppm)</b>	0-0.5	Vinueza-Burgos et al. (2017)
	0.51-0.95	
<b>Conservation temperature (°C)</b>	≤0	Hue et al. (2010)
	>0	
<b>Chilling time (min)</b>	60-100	Hue et al. (2010)
	101-150	
	151-360	
<b>Time in transit to slaughter (h)</b>	<1	
	1-1.5	
	1.6-2	
<b>Carcass weight (kg)</b>	1-2,6	
	2.61-3.2	
	3.21-3.58	

122 ***Statistical analysis***

123 Differences in *Campylobacter* contamination levels at slaughterhouses and processing  
124 stages (after scalding, after de-feathering, after eviscerating, after washing and after  
125 chilling) were assessed by the Chi-square test (at 95% CI and  $p < 0.05$ ).

126 The binary outcome was created to investigate slaughterhouse level factors associated  
127 with the presence of *Campylobacter* (1 if the sample exceeded the criterion of 1,000  
128 CFU/g and 0 if *Campylobacter* enumeration was below 1,000 CFU/g). Multivariable mix-  
129 effect logistic regression was conducted to identify factors associated with  
130 slaughterhouses, reporting the odds ratio (OR) and 95% confidence intervals (CI).  
131 Slaughterhouse was included as a random effect in all analyses. A total of 12 exposure  
132 variables (listed in Table 1) were evaluated as fixed effects in univariable analyses.  
133 Variables with a univariable p-value of  $< 0.10$  were considered for inclusion in a  
134 multivariable model, which was built using a forward stepwise approach. A separate  
135 analysis was also conducted to investigate risk factors associated with *Campylobacter* on  
136 carcasses after chilling. A total of 11 exposure variables (listed in Table 1, with the  
137 exception of processing stages) were evaluated as fixed effects in univariable analyses.  
138 As previously, variables with a univariable p-value of  $< 0.10$  were considered for inclusion  
139 in a multivariable model, which was built using a forward stepwise approach. Models  
140 were run using Stata 13 (College Station, Texas, USA) and the 'xtmelogit' command.

141

142 **RESULTS**

143 ***Samples that exceeded the 1,000 CFU/g criterion during slaughtering***

144 During the study, a total of 1,725 samples were collected from the processing line of 12  
145 different slaughterhouses for *Campylobacter* enumeration. Seventy-five samples from 1  
146 slaughter were discarded due to sampling conditions. Of this amount ( $n=1,725$ ), the

147 23.7% (n = 409) exceeded the 1,000 CFU/g criterion. Statistical differences were shown  
148 in *Campylobacter* counts according to different processing stages (after scalding, after  
149 de-feathering, after eviscerating, after washing and after chilling) and the different  
150 slaughterhouse studied (Figure 2 and 3). Concerning sampling at each processing stage,  
151 the higher counts were shown after the evisceration stage, where 29% of samples  
152 exceeded the limit of 1,000 CFU/g (n = 100/345) and the lowest one after chilling stage  
153 (19.1%, n = 66/345) (Figure 2).

154

155 In terms of critical points, significant differences were found in the processing phases, as  
156 the stages after washing and after chilling showed a decrease in the OR (OR = 0.63,  $P =$   
157 0.031 and OR = 0.58,  $P = 0.013$ , respectively). The total animals slaughtered ranging  
158 from 10,001-20,000 showed an increase in the OR (OR = 2.5,  $P = 0.010$ ). However, those  
159 ranging from 30,001-40,000 presented a decrease in the OR (OR = 0.08,  $P = 0.000$ ).  
160 Moreover, significant differences were found in the flock size, with the 6,501-8,500 range  
161 presenting the decreased OR (OR = 0.04;  $P = 0.000$ ). The age of conventional birds (41-  
162 50 days) was positively associated with a higher *Campylobacter* count (OR = 2.37,  $P =$   
163 0.000). With reference to scalding temperatures, the 51.9-54°C range showed an OR  
164 increase (OR = 3.99,  $P = 0.000$ ). Moreover, *Campylobacter* was isolated more in samples  
165 chilled at temperatures above 0°C (OR = 4.82,  $P = 0.000$ ). In addition, the chilling time  
166 range between 101-150 presented an increased OR (OR = 2.00,  $P = 0.000$ ). Transit time  
167 to slaughter of more than 1 hour presented a significant increase in the OR (1-1.5 h; OR  
168 = 3.20,  $P = 0.000$ ), (1.6-2 h; OR = 13.34,  $P = 0.000$ ) and (>2 h; OR = 5.91,  $P = 0.000$ ).  
169 With respect to the weight of carcasses (3.21-3.58 kg), the range presented a significant  
170 decrease in the OR (OR = 0.32,  $P = 0.003$ ) (Table 3).

171

172 **Table 3.** Final multivariate mixed-effect logistic regression of risk factors associated with  
 173 exceeding the 1,000 CFU/g criterion during all slaughterhouse stages. Slaughterhouse  
 174 was included as random effect.  
 175

Variable	Level	Odds ratio	P-value	95% CI
<b>Processing stages</b>	After scalding	Ref.		
	After de-feathering	0.88	0.547	0.59-1.31
	After eviscerating	1.21	0.328	0.82-1.77
	After washing	0.63	0.031	0.42-0.95
	After chilling	0.58	0.013	0.39-0.89
<b>Number of animals slaughtered (total)</b>	1,000-10,000	Ref.		
	10,001-20,000	2.54	0.010	1.25-5.19
	20,001-30,000	1.66	0.074	0.95-2.89
	30,001-40,000	0.08	0.000	0.03-0.20
	40,001-80,000	0.51	0.226	0.17-1.51
<b>Flock Size (animals)</b>	80,001-100,000	0.71	0.457	0.28-1.74
	500-2,500	Ref.		
	2,501-4,500	0.19	0.000	0.10-0.35
	4,501-6,500	0.24	0.000	0.13-0.44
<b>Slaughter age (days)</b>	6,501-8,550	0.04	0.000	0.01-0.11
	30-40	Ref.		
	41-50	2.37	0.000	1.46-3.85
<b>Scalding temperature (°C)</b>	51-100	1.15	0.747	0.47-2.84
	48-51.8	Ref.		
<b>Conservation temperature (°C)</b>	51.9-54	3.99	0.000	2.59-6.16
	≤0	Ref.		
<b>Chilling time (min)</b>	>0	4.82	0.000	2.87-8.09
	60-100	Ref.		
	101-150	2.00	0.000	1.47-2.71
<b>Time in transit to slaughter (h)</b>	151-360	1.00	0.999	0.67-1.49
	<1	Ref.		
	1-1.5	3.20	0.000	1.81-5.63
	1.6-2	13.34	0.000	6.04-29.44
<b>Carcass weight (kg)</b>	>2	5.91	0.000	2.96-11.81
	1-2,6	Ref.		
	2.61-3.2	0.50	0.000	0.35-0.71
	3.21-3.58	0.32	0.003	0.15-0.68

176 Ref.: Reference. CI: Confident Interval.

177

178 ***Samples that exceeded the 1,000 CFU/g criterion after chilling***

179 Eleven variables were tested and four showed an association with the *Campylobacter* count  
 180 during the univariable analysis after chilling. Findings from the mixed-effects between  
 181 different critical points and the samples that exceeded the criterion of 1,000 CFU/g of  
 182 *Campylobacter* after the chilling stage are summarised in Table 4. Analysis of the flock  
 183 age revealed significantly increased OR in *Campylobacter* enumeration (OR = 7.41, P =

184 0.001). Similarly, scalding temperature (51.9.54 °C) was positively associated with an  
 185 increase in the OR (OR = 2.75,  $P = 0.006$ ). Examining the time in transit at slaughter there  
 186 was a significant OR decrease in samples processed for more than 1-1.5 hours (OR = 0.042,  
 187  $P = 0.000$ ). However, those processed for more than 2 hours presented an increase in the  
 188 OR (OR = 4.44,  $P = 0.050$ ). Regarding carcass weight, the range from 3.21-3.58 presented  
 189 a decrease in the OR (OR = 0.01,  $P = 0.011$ ).

190  
 191 **Table 4.** Final multivariate mixed-effect logistic regression of risk factors associated with  
 192 exceeding the criterion of 1,000 CFU/g after the chilling stage. Slaughterhouse was  
 193 included as random effect.  
 194

Variable	Level	Odds ratio	<i>P</i> -value	95% CI
Slaughter age	30-40	Ref.		
	41-50	7.41	0.001	2.19-25.02
	51-100	3.53	0.205	0.50-25.03
Scalding temperature (°C)	48-51.8	Ref.		
	51.9-54	2.75	0.006	1.68-22.98
Time in transit to slaughter (hours)	<1	Ref.		
	1-1.5	0.25	0.042	0.06-0.95
	1.6-2	0.68	0.687	0.10-4.34
	>2	4.44	0.050	0.99-19.75
Carcass weight (kg)	1-2,6	Ref.		
	2.61-3.2	0.22	0.003	0.08-0.61
	3.21-3.58	0.01	0.011	0.00-0.03

195  
 196 Ref.: Reference. CI: Confident Interval.

197

## 198 DISCUSSION

199 Campylobacteriosis is the main zoonotic gastrointestinal disease in the EU and Spain  
 200 comes fourth in the list of countries reporting the majority of cases (18,860), after  
 201 Germany, the United Kingdom and the Czech Republic [4]. *Campylobacter* colonises the  
 202 poultry gut and the caecal contents of chicken and can contain extremely high  
 203 *Campylobacter* counts ( $10^9$  CFU/g) [17]. Several authors have revealed that the  
 204 evisceration process leads to a considerable increase in cross-contamination of

205 *Campylobacter*, especially as perforation of the intestines often cannot be avoided [18-  
206 20].

207 In this study, mCCDA medium was employed for *Campylobacter* counting, the medium  
208 followed by the standard method [16]. However, it is a selective medium with low  
209 discrimination of real *Campylobacter*, and for this reason the discrimination requires  
210 trained and experienced personnel and time for confirmation tests (microscopic  
211 observation, biochemical testing) [21]. Nevertheless, comparative studies with other  
212 selective media such as CFA (Campy Food Agar), did not reveal statistically significant  
213 differences between *Campylobacter* counts [21-22]

214 Our results showed an increased number of samples that exceeded the criterion of 1,000  
215 CFU/g after evisceration stage, and a 34.5% decrease after the chilling stage. These  
216 results are consistent with others published by Figueroa et al. (2009) and Guerin et al.  
217 (2010), who observed a decrease of 26.6% and 17.5 %, respectively [21-22].

218 In contrast, no statistical differences were found for the batch position (first and last)  
219 within the same slaughter, but statistical differences were observed between the different  
220 slaughterhouses. This suggests that *Campylobacter* presence in the carcasses might be  
221 more associated with the origin of the batch than its position in the slaughter schedule.  
222 Although the entry of positive animals could result in cross-contamination of the  
223 slaughterhouse, it has been reported that only the two subsequent batches are  
224 contaminated in a positive lot [23]. However, the microbiological status of the previous  
225 batch was not considered in the present study.

226

227 After the chilling stage, the following risk factors identified were the age of the animals,  
228 scalding temperature, time in transit to the slaughterhouse and the carcass weight. A  
229 higher percentage of samples exceeded the criterion of 1,000 CFU/g in flocks sacrificed

230 between 41-50 days of age. Lawes et al. (2012) showed an increase in *Campylobacter*  
231 prevalence in animals slaughtered at 46 days of age [5]. Nevertheless, Hue et al. (2010)  
232 showed a high prevalence of *Campylobacter* in older ages (free-range or slow growth)  
233 [24]. Studies suggest that *Campylobacter* broiler colonisation starts at the age of 21 days  
234 (2-3 wks.) and there is a significant increase during the fattening period (from 40 days  
235 onwards) [8].

236

237 Concerning the scalding temperature, our results presented an increase in the OR when  
238 samples were scalded at 51.9-54°C. Procedures for *Campylobacter* control together with  
239 freezing have been shown to reduce the bacteria, although due to the pathogen's ability  
240 to survive in water, in aerosols and on equipment, cross-contamination can occur between  
241 batches, and carcasses free from *Campylobacter* can become contaminated after an  
242 infected batch of birds [23,26]. Regarding time in transit to slaughter, from 1-1.5 hours  
243 was associated with a decrease in *Campylobacter*. However, more than 2 hours tended  
244 to be associated with an increase in the *Campylobacter* presence. This could be due to  
245 the longer time the animals spent in the slaughterhouse, which may be insufficiently  
246 technically equipped and have less machinery (fewer plucking fingers, fewer showers and  
247 fewer evisceration devices). Therefore, more carcasses pass through the same machines,  
248 and more cross-contamination could occur between batches.

249 Regarding carcass weight, although higher levels of *Campylobacter* are reported when  
250 the animals are larger [27], in this case we found a protective factor when the animals  
251 weighed between 3.21-3.58 kg. However, other authors did not find differences between  
252 the body weight and subsequent proliferation of the bacteria [28].

253

254 *Campylobacter* is a confirmed pathogen from gastrointestinal content in poultry.  
255 Measures taken so far have not managed to control the bacteria at field level, as once the  
256 chickens are colonised they remain so until slaughter [31]. Thus, its control has been  
257 regulated at the slaughterhouse. Previous studies reported that reducing *Campylobacter*  
258 prevalence by two logs at field level could lead to a 30-fold reduction in human  
259 campylobacteriosis [31,32]. After that, by following control strategies such as biosecurity  
260 measures at field level together with optimisation of technical and hygienic aspects of the  
261 slaughterhouse, *Campylobacter* counts in chicken could be reduced.

262 Although some studies have reported a prevalence of 70% and 86% in broiler carcasses,  
263 the results of this study suggest that the prevalence that exceeds the limit of 1,000 CFU/g  
264 (limit of infection in humans) is low [5]. Nine of the twelve slaughterhouses analysed in  
265 the study met the criteria set out in the legislation (1,000 CFU/g; c=20; n=50). Direct  
266 comparison of these results to others reported is delicate and should be performed with  
267 caution due to the heterogeneity of the experimental design. To our best knowledge, this  
268 is the first study to examine the risk factor of *Campylobacter* at the slaughterhouse, taking  
269 the EU legislation into account.

270

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