- 1 Fast detection of cocoa shell in cocoa powders by Near Infrared Spectroscopy and
- 2 multivariate analysis

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ABSTRACT

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Cocoa shell must be removed from the cocoa bean before or after the roasting process. In the case of a low efficient peeling process or the intentional addition of cocoa shell to cocoa products (i.e. cocoa powders) to increase the economic benefit, quality of the final product could be unpleasantly affected. In this scenario, the Codex Alimentarius on cocoa and chocolate has established that cocoa cake must not contain more than 5% of cocoa shell and germ (based on fat-free dry matter). Traditional analysis of cocoa shell is very laborious. Thus, the aim of this work is to develop a methodology based on near infrared (NIR) spectroscopy and multivariate analysis for the fast detection of cocoa shell in cocoa powders. For this aim, binary mixtures of cocoa powder and cocoa shell containing increasing proportions of cocoa shell (up to ca. 40% w/w based on fat-free dried matter) have been prepared. After acquiring NIR spectra (1100-2500 nm) of pure samples (cocoa powder and cocoa shell) and mixtures, qualitative and quantitative analysis were done. The qualitative analysis was performed by using principal component analysis (PCA) and partial least squares discriminant analysis (PLS-DA), finding that the model was able to correctly classify all samples containing less than 5% of cocoa shell. The quantitative analysis was performed by using a partial least squares (PLS) regression. The best PLS model was the one constructed using extended multiple signal correction plus orthogonal signal correction pre-treatment using the 6 main wavelengths selected according to the Variable Importance in Projection (VIP) scores. Determination coefficient of prediction and root mean square error of prediction values of 0.967 and 2.43, respectively, confirmed the goodness of the model. According to these results it is possible to conclude that NIR technology in combination with multivariate analysis is a good and fast tool to determine if a cocoa powder contains a cocoa shell content out of Codex Alimentarius specifications.

- 42
- 43 Keywords:
- 44 Cocoa powder
- 45 Cocoa shell
- 46 NIR
- 47 PLS
- 48 PLS-DA

1. Introduction

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Cocoa powder is a cocoa bean (Theobroma cacao) derivative largely consumed around 51 52 the world due to its capacity to give color, flavor and eating pleasure to a myriad of food 53 preparations (Dico et al., 2018). The obtaining of cocoa powder from cocoa beans follows different steps. First of all, 54 55 beans must be peeled, starting with the peeling of the bean before or after a roasting process. During the same peeling, cocoa cotyledon must be separated from cocoa shell 56 (12-20% of the cocoa seed), yielding fragments of cotyledon, called nibs (Okiyama, 57 58 Navarro, & Rodrigues, 2017). During the shelling step, shell should be perfectly separated, removing large parts of shells and leaving nib particles practically unbroken 59 (Beckett, 2009). The performance of this procedure is very relevant since the presence of 60 61 cocoa shell in cocoa beans derivatives (cocoa liquor, cocoa powder or chocolate) adversely affects the final product quality (Mendes & Lima, 2007). Concretely, it can 62 have an influence in some characteristics of the final product such as the flavor or taste; 63 it can also be responsible of off-flavors. Additionally, fiber content in cocoa shell is really 64 high. Thus, it can be a problem for the grinding process, causing equipment abrasion in 65 66 some cases. Bearing this in mind it is not surprising that shell content in cocoa powders is a quality parameter to be controlled. Concretely, the Codex Alimentarius establishes a 67 maximum amount of 5% of cocoa shells in cocoa cake (based on fat-free dry matter) 68 69 (Codex Alimentarius, 2016). 70 Analysis of cocoa shell in cocoa products might be done following the AOAC 968.10 or the 970.23 methods (Codex Alimentarius, 2016). The first method, called spiral vessel 71 72 count consists of counting the spiral vessels in a defatted, grinded and digested sample 73 with the help of a microscope adjusted to mold counting (field of view 1.382 mm at 100x) (AOAC, 1984). The second method, called stone cell count, consists of microscope assisted counting the stone cells present in the samples after a really laborious preparation (AOAC, 1984).

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Since those methods are really arduous, recent attempts to develop alternative methods have been done. Researchers from the Nestlé Research Center proposed a gas-liquid chromatography procedure based on the detection of fatty acid tryptamides (FATs) in the sample, since FATs are compounds more abundant in cocoa shells than in other parts of cocoa seed. This work, carried out with only cocoa originating from the Ivory Coast, demonstrated that it might be an appropriate tool for the determination and prediction of the shell content in cocoa liquor (Hug, Golay, Giuffrida, Dionisi, & Destaillats, 2006). In another work, Yang et al. (2015) proposed the employment of polysaccharide fingerprint established by high performance liquid chromatography followed by principal component analysis to identified cocoa powders adulterated with cocoa or other plant shells such as chestnut, longan, peanut, etc. However, only cocoa powders containing cocoa or other plant shell percentages higher than 15 and 10%, respectively, were detected using this methodology. Therefore, even when these methodologies (determination of FATs, HPLC polysaccharide fingerprint, etc.) are more sensible, accurate and faster than the methods proposed by the Codex Alimentarius, their use as routine techniques for shell content determination still have certain limitations such as the limit of detection or the fact that they need sample preparation, require specialized personnel and they are destructive. To avoid these drawbacks common in traditional chemical analysis techniques, recent attempts on developing accurate and sensitive analytic techniques based on near infrared (NIR) spectroscopy have been done. Due to the ability of NIR spectroscopy to provide a spectrum that acts as a 'fingerprint' distinctive of a particular sample, this technology is now widely used as a successful quality control tool (Lerma-García, Cortés, Talens & Barat, 2018). Concretely, in the cocoa sector NIR spectroscopy has been employed for the prediction of majority (moisture, carbohydrate, fat, protein) or minority functional compounds (theobromine, catechin, organic acids, etc.) (Veselá, Barros, Synytsya, Delgadillo, Čopíková, & Coimbra, 2007; Álvarez et al., 2012; Krähmer et al., 2015) as well as for quality control (discrimination of cocoa beans according to geographical origin, prediction of cocoa powder adulterations, etc) (Teye, Huang, Dai & Chen, 2013; Quelal et al., 2018).

In this scenario, the goal of this work is the fast determination of cocoa shell content in cocoa powders in concentrations higher than the limit established by the Codex Alimentarius (5%) by means of NIR spectroscopy and a multivariate analysis.

2. Materials and methods

2.1. Cocoa powder and shell Samples

A total of 20 natural cocoa powders and 2 cocoa shells, gently provided by Olam Food Ingredients (Cheste, Spain) or purchased in the market from different origins (Ghana, Ivory Coast, Cameroon, Peru and Indonesia) were employed in this study. In order to predict the presence of cocoa shell in cocoa powders using partial least squares (PLS), binary mixtures containing cocoa powder and cocoa shell were prepared. The mixtures contained percentages of cocoa shells in cocoa powder (based on fat-free dry matter) from ca. 2.5 to 40%. Percentages higher than 40% were not considered since over this percentage the presence of cocoa shell is sensory evident. To improve the robustness of the PLS model, all 20 cocoa powder samples (coming from different origins and obtained after different processings) were randomly selected to perform a total of 12 binary

mixtures for each percentage (2.5, 5, 7.5, 10, 20 and 40%), in which both cocoa shell samples were also considered. Thus, a total of 72 mixtures were obtained. Once all mixtures were prepared, they were poured in hermetic plastic containers and stored at 20±2 °C under dark conditions until use.

2.2. NIR spectra acquisition

The 94 samples (20 cocoa powders, 2 cocoa shells and 72 binary mixtures) were measured with a spectrophotometer FOSS NIR 5000 (Silver Spring, MD, USA). A uniform thickness and surface were secured during spectra scanning using a device with 380 mm of diameter and 1cm of thick with a quartz windows which was filled with 5 g of sample. The spectrophotometer gives the measurements in relative absorbance units (log 1/R), which could be correlated with chemical constituents (Liu, Sun, & Ouyang, 2010; Martens, Nielsen, & Engelsen, 2003). Each sample was scanned 32 times in a range comprised between 1100 and 2500 nm at 2 nm intervals (700 points). The samples were measured twice and no differences between them were found.

2.3. Statistical analysis

- Spectral data were pre-treated and analysed using qualitative and quantitative models by means of the chemometric software Unscrambler v10.5 (CAMO Software AS, Oslo, Norway).
- The PCA model was performed using raw data to identify different sample groups and to find and remove defective outliers (Adnan, Hörsten, Pawelzik, & Mörlein, 2017; Bro & Smilde, 2014).
- The PLS was performed in order to predict the presence of cocoa shell in the cocoa

powders and the PLS-DA (Berrueta, Alonso, & Héberger, 2007; Prats-Montalbán, Jerez-149 Rozo, Romañach, & Ferrer, 2012), was constructed to evaluate its capability in 150 classifying samples according to the following categories: cocoa powders containing less 151 152 than 5% cocoa shell (w/w), and cocoa powders containing from 5 to 40% cocoa shell 153 (w/w). 154 Both analyses were performed using the pre-treated spectra. The spectral pretreatments tried included extended multiple signal correction (EMSC) (Martens et al., 155 2003), standard normal variation (SNV), 2nd derivative with the Savitzky-Golay (S-G), 156 orthogonal signal correction (OSC) and combinations of all of them with OSC. 157 158 To construct both PLS and PLS-DA models, two data matrices were used. The first one employed for the PCA and PLS model construction, contained the spectra of all 159 samples (N = 94) and the same 700 X-variables. In this case, all individual cocoa shell 160 161 percentages were considered as Y-variable. The second matrix, employed for the PLS-DA model construction, included the spectra of 92 samples (in which the spectra of cocoa 162 163 shells were not considered since the considered categories were cocoa shell contents 164 below 5% and between 5-40%) and 700 predictors or X-variables (wavelengths), and also a dependent Y-variable containing the 2 categories previously described (<5% and 5-40% 165 166 cocoa shell based on fat-free dry matter, w/w). 167 For both, PLS-DA and PLS models construction, the use of all spectra wavelengths was considered, jointly with the use of the most important wavelengths. The PLS and the 168 score of Variable Importance in Projection (VIP) were combined together for these 169 170 selection (Botelho, Reis, Oliveira, & Sena, 2015). To select the optimal factor number and to avoid the over-fitting of both PLS and PLS-171 172 DA models, leave-one-out cross-validation was used using 70% of the data, which were

randomly selected. The remaining 30% of the data were used as an external validation

174 set.

175 PLS models accuracy was evaluated by the required number of latent variables (LVs), the coefficient of determination of calibration (R²_C), RMSEC, the coefficient of 176 determination of cross-validation (R²_{CV}), RMSECV, the coefficient of determination for 177 prediction (R²_P), the root mean square error of prediction (RMSEP), the ratio of prediction 178 deviation (RPD, which is calculated as ratio between the standard deviation of reference 179 values in training set and RMSEP) and the bias value (which establishes the difference 180 181 between experimental values and NIR predictions). Bias value can be positive (overestimating) or negative (underestimating), indicating values near to zero a minimum 182 183 deviation from experimental and predicted values (Cantor, Hoag, Ellison, Khan, & Lyon, 2011). 184 185 On the other hand, the number of latent variables (LVs) for the PLS-DA model was 186 determined by the low value of the root mean square error of calibration (RMSEC), and 187 the root mean square error of leave-one-out cross validation (RMSECV) (Botelho et al., 188 2015). The PLS-DA classification performance was evaluated by sensitivity, specificity 189 and by the non-error rate (NER). Sensitivity is the model ability related to a correct classification of the samples with different levels of cocoa shell content. The model 190 capacity to correctly determine the samples which not correspond to the class and 191 192 correctly refuse them is the specificity (Almeida, Fidelis, Barata, & Poppi, 2013). The non-error rate (NER) is the average of the sensitivities of the different categories 193 (Manfredi, Robotti, Quasso, Mazzucco, Calabrese, & Marengo, 2018). 194

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3. Results and discussion

3.1. Cocoa powder and shell spectra, pre-treated spectra and PCA analysis

The mean raw spectra of cocoa powders, cocoa shells and binary mixtures of them at different percentages are shown in Fig. 1a. As shown in this figure, the main bands observed appeared at 1470, 1930 and 2130 nm, although other bands at 1730, 2310 and 2350 nm were also evidenced. Although all spectra have a similar pattern of absorbance, the relative absorbance of these bands is different for the different types of samples: cocoa shell is characterized by the highest relative absorbance, which decreased when the content of cocoa shell in the samples decreased. The signal at 1470 nm correspond to the first overtone of O-H and N-H stretching which is associated with a CONH2 structure (peptide) and related to a protein (Osborne, Fearn, & Hindle, 1993). The signal at 1930 nm is related with asymmetric stretching and rocking of water, weakly bounded water, proteins, and aromatics (Veselá et al., 2007), while the wavelength at 2130 nm can be assigned to N-H combination bands (CONH2) (Ribeiro, Ferreira, & Salva, 2011). On the other hand, the band at 1730 nm could be assigned to the first overtone of C-H (Ribeiro et al., 2011), while 2310 and 2350 nm are mostly related to stretching and rocking vibrations of CH2 of polysaccharides (Veselá et al., 2007).

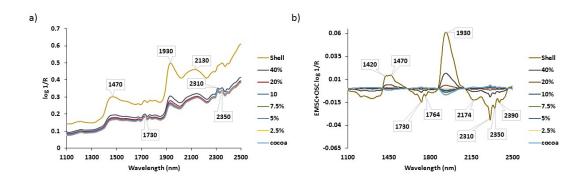


Fig 1. M.A. Quelal-Vásconez et al.

Fig 1. Mean spectra of cocoa powders and shells and mixtures of them at different percentages from (a) raw and (b) pre-treated with EMSC-OSC spectra.

The mean spectra obtained after the application of the EMSC-OSC pre-treatment is shown in Fig. 1b. In this case, the principal wavelengths were 1420, 1470, 1730, 1764, 1930, 2174, 2310, 2350 and 2390 nm. Most of the bands have been previously described, while the other ones could be attributed to the first overtones of symmetric and antisymmetric C-H stretch vibration (CH₂-groups) (1764 nm) (Krähmer et al., 2015), to a combination of C-H (2174 nm) (Ma et al., 2017) and to the combination of C-H stretch and C-H deformation modes (2390 nm) (Wang et al., 2018).

In order to have a more precise idea about the relation between samples and variables a PCA model, a non-supervised method was performed with the raw spectra data to identify possible sample groupings. The score plot of the two first principal components (PCs) is shown in Fig. 2. A total of 98% of the variance is explained by these two first PCs (87 and 11% for PC₁ and PC₂, respectively). Along PC₁, cocoa shell samples were

clearly separated from the remaining ones, in which any clear tendency was observed, although samples containing high cocoa shell percentages (40% w/w) seemed to be located closer to the PC₁ values of cocoa shell. According to the X-loading values (data not shown), the wavelengths with higher discrimination power were 1930, 1420 and 1470 nm for the PC₁ and for the PC₂ were 1644, 1326, 2146, 2310 and 2350 nm. Some of these peaks (1930, 1470, 2310 and 2350 nm) matched with the main peaks observed in raw spectra, which have been previously mentioned. The other bands corresponded to the fist overtone of the hydroxyl and amino groups (1420 nm) and first overtone of C-H (1644 nm) (Ribeiro et al., 2011), the second overtone of C-H (1326 nm) (Ma, Wang, Chen, Cheng, & Lai, 2017) and the combination of C-C and C-H stretching (2146 nm) (Workman, & Weyer, 2008).

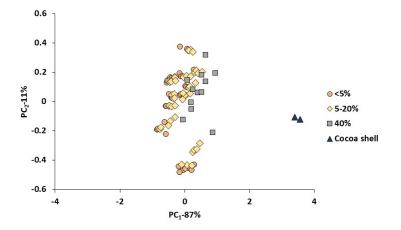


Fig 2. M.A. Quelal-Vásconez et al.

Fig 2. PCA score plot of the two first PCs showing the distribution of all the samples considered in this study. Samples were labelled as follows: cocoa shell content < 5%, comprised between 5 and 20%, 40% and pure cocoa shells.

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A total of 8 PLS models using all the available wavelengths (700) as variables, one for each pre-treatment considered in the study, were performed. The results obtained are summarized in Table 1. At the sight of the results, the best PLS model was the one constructed using the EMSC+OSC pre-treatment. In order to reduce the high dimensionality of the spectral data, the most important wavelengths were selected according to the VIP scores (figure 3). These VIP scores determine the significance of each variable in the projection used by a given PLS model by means of their coefficients in every component, jointly with the significance of each component in regression (Botelho et al., 2015). As it could be observed in Fig. 3, the most important variables are wavelengths at 1930, 1420 and 1470 nm at positive values of LV₁, and 2310, 2350 and 1730 nm at negative values of LV₁. These wavelengths are mostly the same previously mentioned in both raw and pre-treated spectra, which demonstrated their importance in cocoa shell content prediction. Most of these wavelengths have been previously described in literature in the prediction of several compounds (such as fat, carbohydrates, polysaccharides, moisture, polyphenols, etc.) of cocoa beans and derived products (Huang et al., 2014; Krähmer et al., 2015; Quelal-Vásconez et al., 2018; Veselá et al., 2007). Using the EMSC+OSC pre-treatment and the six wavelengths obtained in the VIP scores as variables, another PLS model was constructed. The results obtained for this model are also shown in Table 1. Compared to the best model obtained with the same pre-treatment but using all the available wavelengths, this model is less complex although all the other parameter values are very similar.

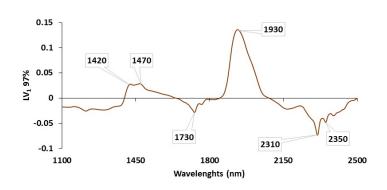


Fig. 3. M.A. Quelal-Vásconez et al.

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Fig 3. Variable importance in projection (VIP) scores of the PLS model constructed to predict cocoa shell percentages.

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Table 1

Results of the PLS models constructed for predicting cocoa shell percentage using different pre-treatments and different number of wavelengths with a calibration and validation sets.

Pretreatment	#W	#LV	Calibration		Cross-validation		Validation			
			$\mathbf{R^2}_{\mathbf{C}}$	RMSEC	R ² cv	RMSECV	\mathbb{R}^2 P	RMSEP	Bias	RPD
Raw data	700	7	0.908	3.68	0.694	6.83	0.930	3.52	0.351	3.46
EMSC	700	7	0.936	3.06	0.857	4.64	0.941	3.24	0.095	3.77
SNV	700	7	0.931	3.18	0,862	4.55	0.940	3.27	0.057	3.72
2nd Der. (S-G)	700	7	0.967	2.20	0.936	3.09	0.955	2.96	-0.021	4.11
OSC	700	1	0.990	1.20	0.989	1.25	0.851	5.16	-0.059	2.36
EMSC-OSC	700	1	0.974	1.92	0.973	2.01	0.967	2.41	0.204	5.06
SNV+OSC	700	1	0.978	1.79	0.976	1.89	0.967	2.55	-0.278	4.77
2nd Der. (S-G)+OSC	700	3	0.944	2.85	0.942	2.96	0.939	3.33	-0.104	3.66
EMSC-OSC	6	1	0.975	1.91	0.973	2.01	0.967	2.43	0.195	5.03

#W = number of wavelengths used to construct de model; #LV = latent variables; R^2 =

determination coefficient; RMSEC = Root mean square error of calibration; RMSECV =

Root mean square error of cross-validation; RMSEP = Root mean square error of

prediction; RPD = Ratio prediction deviation; EMSC = Extended multiple scatter correction; 2nd Der. (S-G) = Second derivative and Savitzky Golay smoothing, SNV = Standard Normal Variate, OSC = Orthogonal signal correction.

The plot representing the predicted versus the measured cocoa shell percentages of the prediction set samples constructed with PLS data of the model constructed using the 6 wavelengths as variables is shown in Fig. 4. A good linear fit due to the closer relationship between the reference values and the NIR spectra is observed, displaying the reliability and accuracy of the NIR in determining the percentage of cocoa shell present in the cocoa powders.

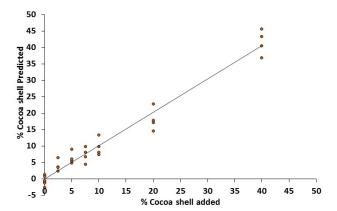


Fig. 4. M.A. Quelal-Vásconez et al.

Fig 4. Predicted versus measured cocoa shell percentages by PLS model constructed using the 6 main wavelengths in the prediction set.

3.3. Classification of cocoa powder samples according to the added level of cocoa shell

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Since PCA is a non-supervised method, and it is not possible to observe a clear separation between the different sample categories, a supervised discriminant model, PLS-DA, was next constructed using all the available wavelengths (700) and the EMSC-OSC pre-treatment. The best model was obtained with 2 LVs with RMSEC and RMSECV values of 0.24 and 0.28, respectively, with most of the variability explained by the LV₁ (72%). Next, using the 6 most relevant wavelengths as variables, another PLS-DA model was constructed. The discriminant plot obtained using the two LVs for the classification of samples according to the different categories is shown in Fig. 5. As it can be observed in this figure, separation between the two categories is achieved along LV_1 , with negative scores related to the samples containing < 5% cocoa shell, and positive scores related to samples containing 5-40% cocoa shell. Once constructed, the model was validated with the external validation set samples. The results obtained for both calibration and external validation sets for this model are included in Table 2. As it can be observed in the confusion table for the calibration samples, all samples were correctly classified. On the other hand, for the external validation set, all samples of the <5% category were correctly classified, while 3 samples of the 5-40% category were misclassified. Even if the number of misclassified samples is very low, it should be highlighted that all the "misclassified samples" corresponded to samples containing a 5% cocoa shell (based on fat-free dry matter), which is the limit established by the Codex Alimentarius, and thus the borderline of both categories. Next, the PLS-DA classification performance was evaluated by the sensitivity, specificity and NER values, which are also included in Table 2. Taking into account the values reported and the comments previously mentioned, it could be concluded that the PLS-DA model constructed is able to reliable discriminate between samples containing cocoa shell percentages below and upper 5%.

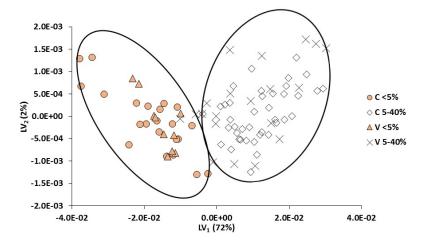


Fig. 5. M.A. Quelal-Vásconez et al.

Fig 5. PLS-DA discriminant plot constructed using the two first LVs of the model constructed using the 6 main wavelengths to classify cocoa powders according to the following categories: cocoa shell content < 5% and cocoa shell content comprised between 5 and 40%. Both calibration (C<5% and C 5-40%) and external validation (V<5% and V 5-40%) set samples have been included and represented with different symbols.

Table 2

Confusion table, sensitivity (SENS), specificity (SPEC) and non-error prediction rates (NER) of the PLS-DA model constructed with variable selection to discriminate cocoa powders into two categories: cocoa powders with < 5% and between 5-40% cocoa shell.

Calibration set samples

	Category		# Samples				
	<5%	5-40%		SENS (%)	SPEC (%)	NER (%)	
<5%	22	0	22	100	100	100	
5-40%	0	40	40	100	100	100	
	22	40	62				

External validation set samples

	Category		Category # Samples				
	<5%	5-40%		SENS (%)	SPEC (%)	NER (%)	
<5%	10	0	10	100	85	02.5	
5-40%	3	17	20	85	100	92.5	
	13	17	30				

4. Conclusions

NIR spectroscopy in combination with PLS and PLS-DA statistical models has been shown to be a rapid and effective method to determine cocoa shell content in cocoa powders. Using a PLS analysis, it was possible to quantify the percentage of cocoa shell present in cocoa powders. The best PLS prediction model was constructed using the 6 main wavelengths (1420, 1470, 1730, 1930, 2310 and 2350) selected according to the VIP scores, obtaining 1 LV with R²_C and R²_{CV} of 0.975 and 0.973, respectively, and RMSEC and RMSECV of 1.91 and 2.01, respectively. Regarding the validation samples, R²_P was 0.967 while RMSEP was 2.43, confirming the goodness of the model. On the

other hand, the PLS-DA analysis show that 92.5% of the validation set samples were correctly classified into two groups: samples with a shell content lower than 5% (considered the acceptance limit in cocoa powders by the Codex Alimentarius) and shell contents between 5 and 40%. These results indicate that this technology is therefore an important tool for cocoa producers and clients, who will be able to discriminate among samples in or out specifications, avoiding the use of destructive techniques that require a complex preparation of the sample or techniques that imply an important expense for the company.

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