

Role of lipoprotein lipase activity on lipoprotein metabolism and the fate of circulating triglycerides in pregnancy

Emilio Herrera, PhD, Miguel A. Lasunción, PhD, Diego Gomez-Coronado, PhD,
P. Aranda, PhD, Pilar López-Luna, PhD, and Isabel Maier, MsSc

Madrid, Spain

The mechanism that induces maternal hypertriglyceridemia in late normal pregnancy, and its physiologic significance are reviewed as a model of the effects of sex steroids on lipoprotein metabolism. In the pregnant rat, maternal carcass fat content progressively increases up to day 19 of gestation, then declines at day 21. The decline may be explained by the augmented lipolytic activity in adipose tissue that is seen in late pregnancy in the rat. This change causes maternal circulating free fatty acids and glycerol levels to rise. Although the liver is the main receptor organ for these metabolites, liver triglyceride content is reduced. Circulating triglycerides and very-low-density lipoprotein (VLDL)-triglyceride levels are highly augmented in the pregnant rat, indicating that liver-synthesized triglycerides are rapidly released into the circulation. Similar increments in circulating VLDL-triglycerides are seen in pregnant women during the third trimester of gestation. This increase is coincident with a decrease in plasma postheparin lipoprotein lipase activity, indicating a reduced removal of circulating triglycerides by maternal tissues or a redistribution in their use among the different tissues. During late gestation in the rat, tissue lipoprotein lipase activity varies in different directions; it decreases in adipose tissue, the liver, and to a smaller extent the heart, but increases in placental and mammary gland tissue. These changes play an important role in the fate of circulating triglycerides, which are diverted from uptake by adipose tissue to uptake by the mammary gland for milk synthesis, and probably by the placenta for hydrolysis and transfer of released nonesterified fatty acids to the fetus. After 24 hours of starvation, lipoprotein lipase activity in the liver greatly increases in the rat in late pregnancy; this change is not seen in virgin animals. This alteration is similar to that seen in liver triglyceride content and plasma ketone body concentration in the fasted pregnant rat. In the fasting condition during late gestation, heightened lipoprotein lipase activity is the proposed mechanism through which the liver becomes an acceptor of circulating triglycerides, allowing their use as ketogenic substrates, so that both maternal and fetal tissues may indirectly benefit from maternal hypertriglyceridemia. Changes in the magnitude and direction of lipoprotein lipase activity in different tissues during gestation actively contribute both to the development of hypertriglyceridemia and to the metabolic fate of circulating triglycerides. Any deviation in these metabolic adaptations occurring in the human mother may have consequences that modify her lipoprotein profile, even postpartum. Hormone-induced changes in pregnancy mirror those seen with oral contraceptive steroids and provide a teleologic rationale for the lipoprotein changes induced by sex steroids. (AM J OBSTET GYNECOL 1988;158:1575-83.)

Key words: Hypertriglyceridemia, lipoprotein lipase, lipoproteins, pregnancy

Maternal hypertriglyceridemia is one of the most striking and consistent changes occurring at late gestation in both humans¹⁻⁴ and experimental animals,⁵⁻⁷ but its physiologic significance is not yet completely understood. The fetus does not directly benefit from this maternal condition. The passage of intact triacylglycerol across the placenta has not been detected in any study.⁸ It is known that the supraphysiologic increase in circulating triglyceride-rich lipoproteins dur-

ing pregnancy constitutes a risk of permanent hyperlipidemia postpartum.⁹ We review major adaptations of triglyceride metabolism occurring during late gestation to attain a better understanding of the mechanism for the induction of maternal hypertriglyceridemia in normal pregnancy and the physiologic significance of this change. The key role of maternal lipoprotein lipase activity in determining the tissular fate of circulating triglycerides is emphasized. For ethical reasons, most data were derived from studies of the pregnant rat, but some data from pregnant women are also presented to document the validity of the extrapolation from the experimental data.

Material and methods

Studies in the rat. Female Wistar rats from our own colony and weighing 160 to 180 gm were mated at 2

From the Departamento de Bioquímica, Hospital Ramon y Cajal and Universidad de Alcalá de Henares.

Supported by a grant from the U.S.-Spain Joint Committee for Scientific and Technological Cooperation.

Reprint requests: Emilio Herrera, PhD, Servicio de Bioquímica, Hospital Ramon y Cajal, Ctra. Colmenar Km 9, 28034-Madrid, Spain.

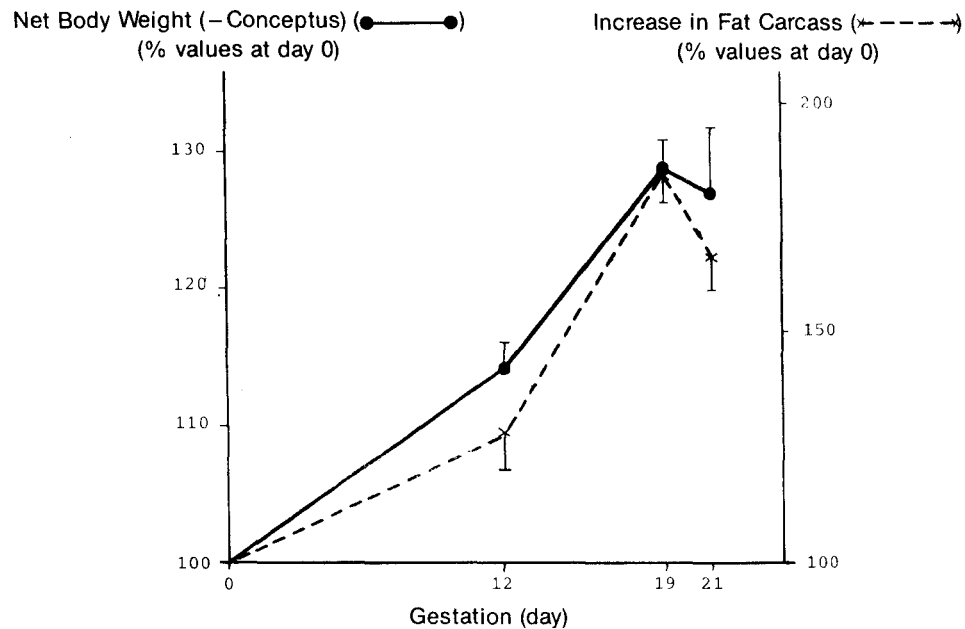


Fig. 1. Change of maternal free-conceptus body weight (solid line) and carcass fat content (dashed line) in the pregnant rat. $n =$ Five to seven rats per group.

Table I. Plasma triglycerides and VLDL lipids in 20-day-pregnant and nonpregnant rats

	Nonpregnant	Pregnant	p
Triglycerides (gm/dl)	76.5 \pm 5.6	390.4 \pm 61.3	<0.001
VLDL-triglycerides (mg/dl)	31.6 \pm 2.5	232.7 \pm 25.3	<0.001
VLDL-cholesterol (mg/dl)	2.5 \pm 0.1	13.9 \pm 0.3	<0.001
VLDL-triglycerides/VLDL-cholesterol (mg/dl)	12.1 \pm 0.4	15.8 \pm 0.3	<0.001

$n =$ Four to seven rats per group. $p =$ Significance of the difference between pregnant and nonpregnant rats.

months of age. Gestational time was estimated by the appearance of spermatozooids in vaginal smears. Age- and sex-matched virgin animals were always studied in parallel with the experimental animals. All animals were killed by decapitation, and blood collected from the neck wound was processed for plasma metabolite analysis and isolation of lipoproteins by sequential ultracentrifugation.⁷⁻¹⁰ For the analysis of very-low-density lipoprotein (VLDL) subfraction distribution, purified VLDL, corresponding to 1.8 mg triglycerides, was applied to heparin-Sephacrose CL 6B columns in 5 mmol/L Tris HCl buffer (pH 7.4) containing 0.05 mol/L NaCl and 0.025 mmol/L MnCl₂ and eluted stepwise with 0.05, 0.12, 0.20, 0.50, and 1.00 mol/L NaCl in 5 mmol/L Tris HCl buffer. Carcass fat content was determined by both body specific gravity and gravimetry of purified lipid extracts.¹¹ Adipose tissue lipolysis was determined in lumbar fat pad pieces incubated in vitro in the presence of U-¹⁴C-glycerol (0.5 μ Ci/ml), fatty acid-free bovine albumin (10 mg/ml), and glu-

cose (5 mmol/L), as described elsewhere.^{12,13} Liver triglycerides were measured in lipid extracts after removal of phospholipids.¹⁰ For determination of lipoprotein lipase activity, tissues were rapidly excised and placed in liquid nitrogen, and the enzyme was assayed in acetone/diethyl ether extracts as previously described.¹¹ Values were expressed as a function of protein concentration.¹⁵

Studies in pregnant women. Twelve pregnant women who registered for obstetric care were studied. Subjects were selected according to the following criteria: (1) values from routine analysis in normal range for gestational period of at least three blood samples obtained during gestation, (2) normal course and outcome of pregnancy, (3) no endocrine abnormalities, (4) single pregnancy, and (5) customary diets and no medication except multivitamin and iron supplementation. All women gave informed consent and were seen during the first and third trimesters of gestation. Blood samples were obtained in tubes containing 1.5 mg/ml

ethylenediaminetetraacetic acid (EDTA) from seated subjects after an overnight fast. After centrifugation, plasma was subjected to ultracentrifugation at $d = 1.006$ for VLDL purification and analysis of VLDL-triacylglycerol and VLDL-cholesterol (VLDL-C) in a Hitachi 705 automatic analyzer (Hitachi, Tokyo, Japan). Aliquots of VLDL were used for studying heparin-Sepharose profiles, as indicated above. After the blood samples were taken for lipoprotein analysis, heparin was injected intravenously in a dose of 50 IU/kg. Ten minutes after heparin injection a second blood sample was taken into a heparinized tube kept on ice. Plasma was separated and stored at -80°C until assayed. Postheparin lipoprotein lipase was measured by using a stable and specific substrate,^{16,17} and potential interference of hepatic lipase in the assay was eliminated by assaying parallel samples in the presence of 1.0 mol/L NaCl.

Expression of results. Results are expressed as mean \pm SEM; statistical comparison between groups was done by Student *t* test.

Results and comment

In normal pregnancy maternal fat storage occurs during the early stages of gestation; in later phases an accelerated transfer of glucose and amino acids to the fetus occurs. As shown in Fig. 1, the maternal, conceptus-free body weight of the pregnant rat progressively increases up to day 19 of gestation and then stops; this change is paralleled by the increase in maternal carcass fat content, which clearly declines at day 21. During late gestation an intense extraction of maternal glucose and amino acids by the placenta occurs to support the rapid fetal accretion,^{18,21} and these changes are responsible for the maternal tendency toward hypoglycemia.²²

Heightened glucose and amino acid extraction and the endocrine changes occurring in the mother have intense effects on the metabolic activity of adipose tissue. As shown in Fig. 2, in vitro lipolytic activity in pieces of lumbar fat pads from 19-day-pregnant rats is higher than activity in controls. This difference is enhanced under fasting conditions, in which greater maternal hypoglycemia²³ and reactive endocrine changes, like enhanced catecholamine production,²⁴ stimulate maximal breakdown of maternal fat depots.

The active adipose tissue lipolytic activity during late gestation is responsible for both the arrest in the progressive increment of fat storage seen during earlier phases (Fig. 1) and, as shown in Fig. 3, for the increase of free fatty acid and glycerol levels found in maternal circulation. The liver is the main receptor organ for these lipolytic products,²⁵ and may either metabolize them through different pathways or reesterify them

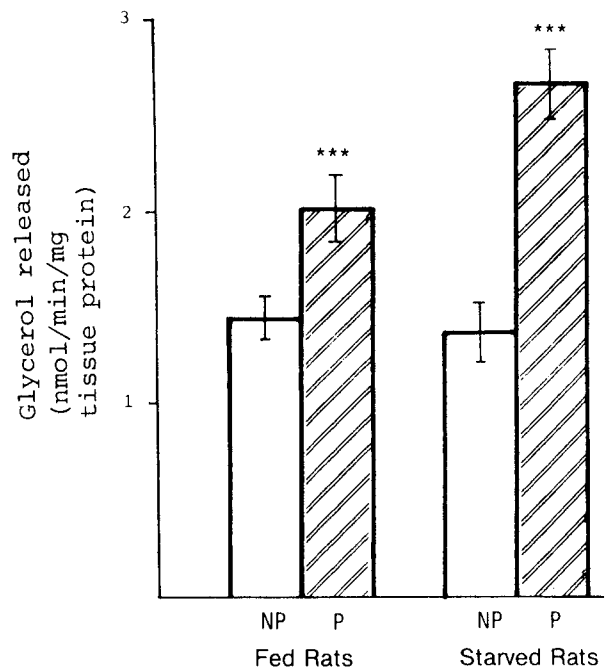


Fig. 2. Rates of in vitro lipolysis (glycerol released) in pieces of lumbar fat pads from fed and 48-hour fasted 19-day-pregnant rats (*P*) and their nonpregnant controls (*NP*). Asterisks correspond to the significance of the difference between pregnant and nonpregnant rats ($***p < 0.001$). $n =$ Six rats per group.

for triglyceride synthesis and later release to circulation in the form of VLDL.

Triglyceride liver content in the rat mother is decreased during late gestation, but circulating triglycerides are greatly enhanced (Fig. 3). These findings indicate that triglycerides synthesized in liver from circulating free fatty acids and glycerol or from endogenous substrates are rapidly released into the circulation. In agreement with this interpretation, it can be seen in Table I that maternal hypertriglyceridemia in the late pregnant rat specifically corresponds to an increase in circulating VLDL-triglycerides, and although VLDL-cholesterol levels are also enhanced, the triglyceride/cholesterol ratio in VLDLs is significantly greater in pregnant than in nonpregnant rats. An enhanced production of triglyceride-rich lipoproteins (presumably chylomicrons) by the intestine from dietary lipids is also seen in late pregnancy in the rat, as indicated by the quicker and greater appearance of labeled lipids in plasma shortly after oral administration of ^{14}C -tripalmitine in 20- and 21-day-pregnant rats compared with controls.²⁶ These findings are also consistent with the increased production of triglyceride-rich lipoproteins in the late pregnant rat, as previously suggested by Childs et al.²⁷

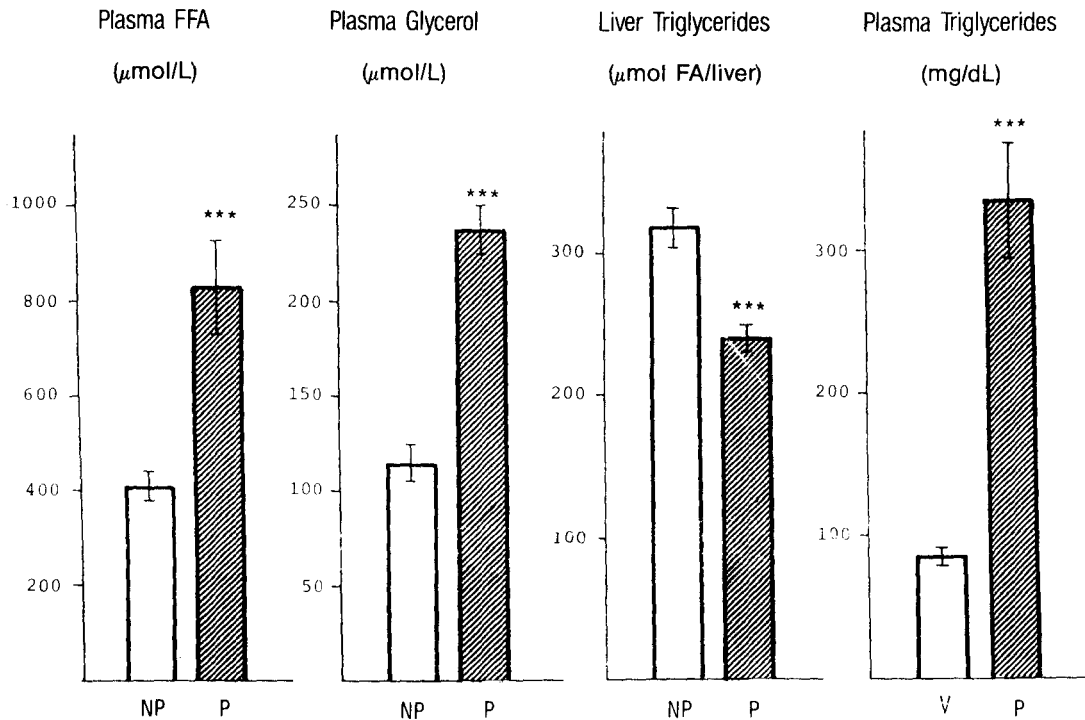


Fig. 3. Plasma metabolite levels and liver triglyceride content in 20-day-pregnant rats (*P*) and their nonpregnant controls (*NP*). Asterisks correspond to the significance of the difference between pregnant and nonpregnant rats (***) $p < 0.001$. $n =$ Six to eight rats per group.

Table II. Plasma VLDL lipids and postheparin lipoprotein lipase activity in pregnant women

	First trimester	Third trimester	<i>p</i>
VLDL-triglycerides (mg/dl)	31.0 ± 5.1	112.8 ± 16.5	<0.001
VLDL-cholesterol (mg/dl)	5.22 ± 0.85	21.92 ± 3.13	<0.001
VLDL-triglycerides/VLDL-cholesterol (mg/dl)	5.83 ± 0.23	5.28 ± 0.35	NS
Postheparin lipoprotein activity (nkat/ml)	424 ± 51	71 ± 16	<0.001

$n = 12$ women per group. $p =$ Significance of the difference between third and first trimesters of gestation.

As shown in Table II, the increase in circulating VLDL-triglycerides and VLDL-cholesterol also occurs in women during late gestation. However, lipoprotein metabolism differs from that in the rat, because the VLDL-triglycerides/VLDL-cholesterol ratio remains unchanged. Although the reason for this species difference is not yet known, it may be caused by the lack of neutral lipid transfer protein in the rat^{28,29} that is, however, present in humans.³⁰ This lipid transfer protein is known to mediate the exchange of esterified cholesterol and triglycerides between VLDL and higher density lipoproteins. The absence of this protein in the rat may result in the specific accumulation of triglycerides in the VLDL particles, whereas in women they are more easily transferred to lipopro-

teins of higher density. This interpretation agrees with our unpublished findings of an enhanced triglyceride/cholesterol ratio in both low-density lipoprotein (LDL) and high-density lipoprotein (HDL) in women in the third trimester of pregnancy. The unchanged VLDL-triglycerides/VLDL-cholesterol ratio in pregnant women also agrees with the fact that the proportional distribution of VLDL subfractions does not change in human pregnancy, as indicated by the heparin-Sepharose elution profile in Fig. 4. We had previously found that this profile was greatly modified in late pregnancy in the rat.³¹

The benefits to the mother from the increase in circulating triglycerides is not fully understood. Circulating maternal triglycerides cannot be directly used to

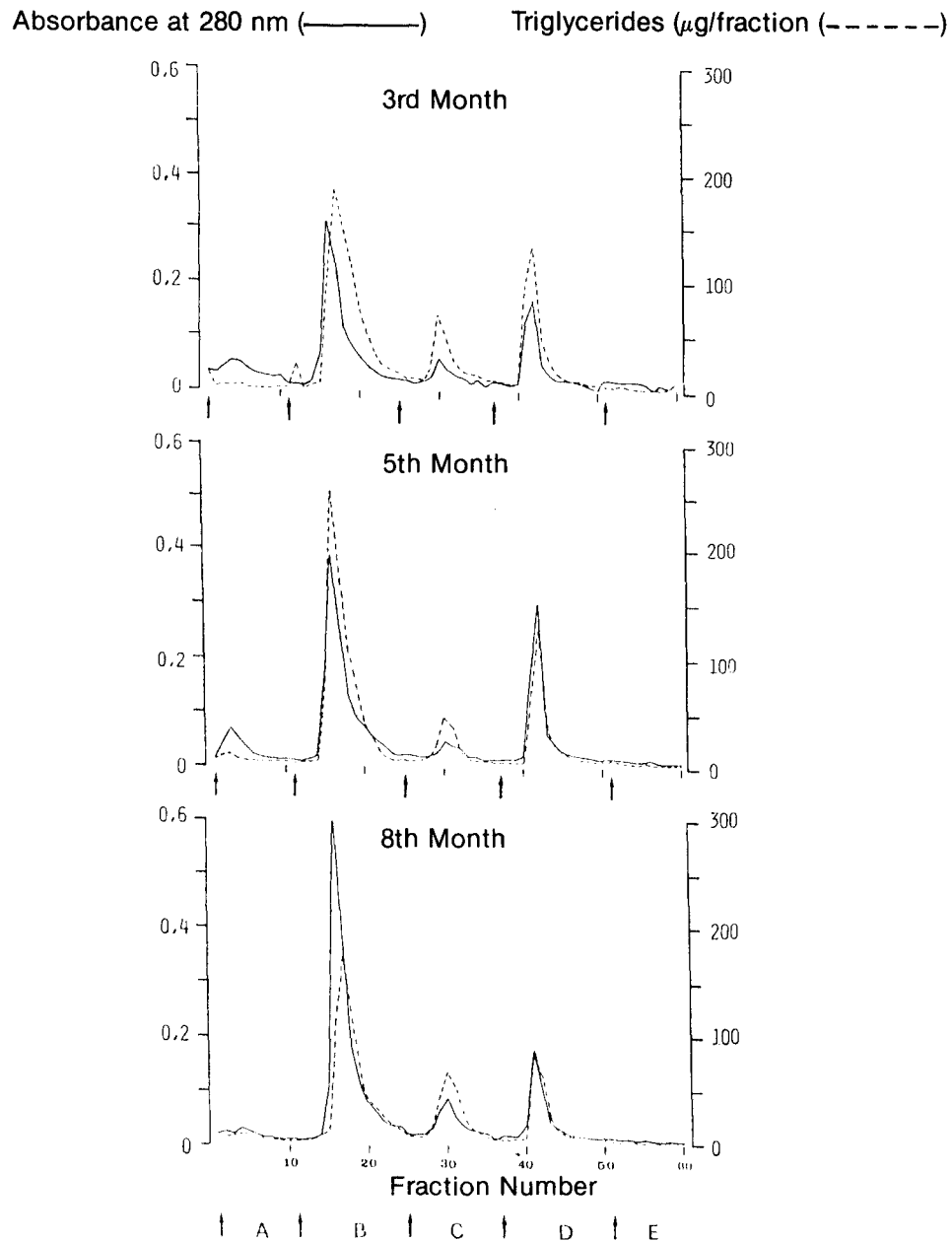


Fig. 4. Heparin-Sepharose column elution profile in 5 mmol/L Tris buffer, pH 7.4, supplemented with 0.05 (A), 0.12 (B), 0.20 (C), 0.50 (D), or 1.00 (E) mol/L NaCl of VLDL ($d < 1.006$) purified by ultracentrifugation from plasma of pregnant women at different months of gestation. Values of absorbance at 280 nm are indicated by solid lines, and those of triglyceride content per fraction (μg) by dashed lines.

support fetal metabolic needs, because they cannot cross the placental barrier.⁸ We have focused our attention on maternal lipoprotein lipase activity, because this enzyme catalyzes triglyceride hydrolysis in circulating triglyceride-rich lipoproteins³² and facilitates hydrolytic products uptake by the subjacent tissue.³³ Because this enzyme is normally bound to capillary en-

dothelium and is released by heparin,³¹ we measured its activity in plasma from pregnant women 10 minutes after administration of 50 IU/kg intravenous heparin. As shown in Table II, postheparin lipoprotein lipase activity decreases at late gestation. This change exceeds the extent to which lipoprotein lipase would be diluted in a larger plasma volume and occurs when maternal

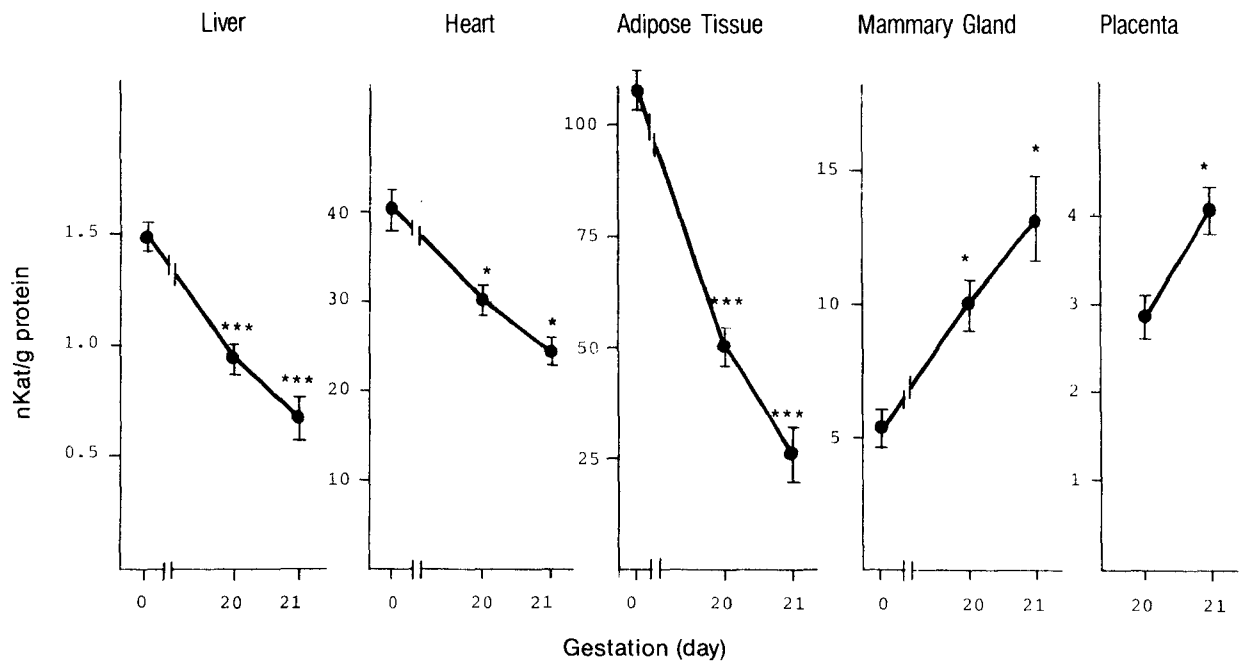


Fig. 5. Effect of pregnancy on lipoprotein lipase activity in different rat tissues. Statistical comparisons versus values in nonpregnant animals (day 0) are shown by asterisks (* $p < 0.005$, *** $p < 0.001$). In the case of the placenta, the asterisk corresponds to the comparison between 21 and 20 days of gestation. $n =$ Six to eight rats per group.

circulating triglycerides are highest. Our findings indicate either that the overall plasma triglyceride removal capacity by maternal tissues decreases as gestation advances or that the different tissues redistribute their use of circulating triglycerides. To determine how specific tissues participate in this reduced postheparin lipoprotein lipase activity, lipoprotein lipase activity was measured in different tissues in the pregnant rat. As shown in Fig. 5, and as was expected, the highest activity in nonpregnant rats was found in adipose tissue, and the lowest in the liver. During late gestation a significant decline in lipoprotein lipase activity was observed in liver, heart, and adipose tissue, whereas an increase was seen in mammary gland and placenta between days 20 and 21 of gestation. In keeping with the decline in fat stores in late gestation (Fig. 1) and with the reductions in liver triglycerides (Fig. 3), these findings suggest that circulating triglycerides, whether alimentary or endogenous, are diverted from those tissues where lipoprotein lipase activity decreases (liver, heart, adipose tissue) and are taken up by the mammary gland, placenta, and possibly other maternal tissues in which enzyme activity increases.

Changes in lipoprotein lipase activity in specific tissues may have important metabolic implications for both the mother and the fetus. The intense reduction of lipoprotein lipase activity found in maternal adipose tissue during late gestation confirms previous reports

from this^{10,14} and other laboratories.^{35,36} Because of the high level of lipoprotein lipase activity present in this tissue as compared with others under nonpregnant conditions (Fig. 5), and because of its high proportional corporeal mass, the reduction in adipose tissue lipoprotein lipase activity must produce a reduction in the clearance of circulating triglyceride-rich lipoproteins. This hypothesis is supported by the diminished uptake of VLDL-triglycerides in isolated adipocytes taken from rats during late pregnancy³⁷; this effect and the previously described increase in circulating triacylglycerol production constitute the major factors responsible for the intense maternal hypertriglyceridemia observed during late gestation.

Progressive increments in placental lipoprotein lipase activity both facilitate the hydrolysis of maternal triglycerides and direct the transfer of the released non-esterified fatty acids across the tissue to the fetus, as has already been proposed to occur in different species, including humans.³⁸⁻⁴⁰

The low lipoprotein lipase activity in maternal liver may contribute to the maintenance of a low concentration of triglycerides in that organ in the fed state. Inasmuch as we had previously found that lipoprotein lipase activity increased in the liver of newborn rats under fasting conditions and that this change was related to high ketone body production,⁴¹ lipoprotein lipase activity was measured also in the fasted pregnant

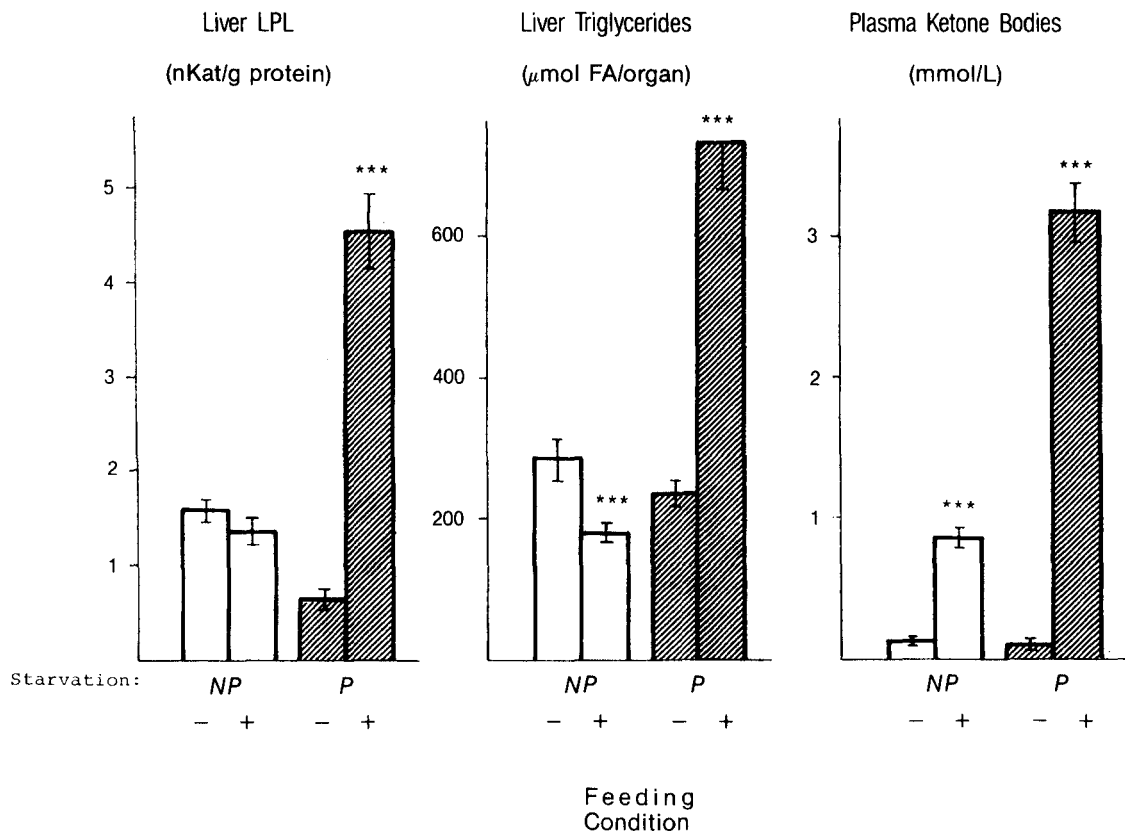


Fig. 6. Effect of 24-hour starvation on liver lipoprotein lipase activity and triglyceride content and plasma ketone bodies in the 20-day-pregnant rats (*P*) and their nonpregnant controls (*NP*). Statistical comparison between fed (+) and starved (-) groups are shown by asterisks (** $p < 0.01$, *** $p < 0.001$). $n =$ Six to eight rats per group.

rat, in which ketone body production is greatly enhanced as well.^{23, 42} As shown in Fig. 6, although 24-hour starvation does not affect liver lipoprotein lipase activity in nonpregnant rats, it causes a marked increase in lipoprotein lipase activity in pregnant rats. A parallel picture is seen in both liver triglyceride and plasma ketone body concentrations. In the pregnant rat, after 24-hour starvation, concentration of these substances increases greatly, whereas a much smaller increment is observed in the nonpregnant rat (Fig. 6). This finding points to the interconnection of these three parameters. Data in the literature show that in the nonpregnant, adult, fasted rat liver, lipoprotein lipase activity may be enhanced by the administration of intralipid, suggesting a substrate inductor mechanism.⁴³

It is not yet known whether the observed liver lipoprotein lipase increment in the fasted pregnant rat has an intrahepatic or extrahepatic origin, but the exaggerated hypertriglyceridemia observed during pregnancy may be proposed as the inductor process. Through this mechanism the liver, which under normal conditions is a triglyceride-exporter organ, becomes a

heightened acceptor of circulating triglycerides, thus allowing their increased consumption as ketogenic substrates. Enhanced production of ketone bodies in the fasted state benefits both fetal and maternal metabolism, because the availability of other substrates is limited. Ketone bodies easily cross the placental barrier, and may be used as energy sources by the fetus^{44, 45}; in the mother, ketone body metabolism may compensate for hypoglycemia, particularly in maintaining cerebral oxidative metabolism, as well as for the reduced utilization of circulating triglycerides caused by low lipoprotein lipase activity.

The other maternal tissue in which lipoprotein lipase activity is greatly modified during late gestation is the mammary gland (Fig. 5). In the rat model it was previously found that just prior to parturition maternal hypertriglyceridemia declined and approached pre-gestation levels.¹⁰ Although not tested around parturition, a similar change has been described in pregnant women after delivery.^{4, 9, 16} This effect occurred in the rat with no change in adipose tissue lipoprotein lipase activity, the values of which remained very low at this

time, but it did coincide with a further increase in mammary gland lipoprotein lipase activity.¹⁰ The direct relationship between these two changes is supported by our previously reported findings.¹⁰ In the pregnant rat that reduction in circulating triglycerides did not occur when the induction of the enzyme in the mammary gland was blocked by progesterone treatment, which avoids the prolactin peak normally occurring before parturition.¹⁰ We therefore conclude that, around parturition, the increase in circulating triglyceride-rich lipoproteins is not as high as would be expected, or a decline may even occur, because of augmented removal through mammary gland lipoprotein lipase activity. In this way maternal circulating triglycerides are diverted to milk synthesis in preparation for lactation.

We conclude that changes in the magnitude and even the direction of lipoprotein lipase activity in different tissues during gestation actively contribute not only to the development of hypertriglyceridemia but also to the metabolic fate of circulating triglycerides. Through this mechanism several needs of maternal and fetal metabolism are fulfilled. However, any change in these dynamic and intricate metabolic adaptations seen in the mother throughout gestation may directly modify her lipoprotein profile. Under pathologic conditions the alteration may be permanently maintained, thereby increasing the risk for the development of cardiovascular disease. Hormone-induced changes in pregnancy mirror those seen with oral contraceptive steroids and provide a teleologic rationale for the lipoprotein changes induced by sex steroids.

We thank Carol F. Warren for editorial help and M. A. Murua for technical assistance.

REFERENCES

- Knopp RH, Montes A, Warth MR. Carbohydrate and lipid metabolism during pregnancy. In: Laboratory indices of nutritional status in pregnancy. Washington, DC: National Academy of Sciences, 1978:35-88.
- Potter JM, Nestel PJ. The hyperlipidemia of pregnancy in normal and complicated pregnancies. *AM J OBSTET GYNECOL* 1979;133:165-70.
- Fahraeus L, Larsson-Cohn U, Wallentin L. Plasma lipoproteins including high density lipoprotein subfractions during normal pregnancy. *Obstet Gynecol* 1985;66:468-72.
- Knopp RH, Warth MR, Charles D, et al. Lipoprotein metabolism in pregnancy, fat transport to the fetus, and the effects of diabetes. *Biol Neonate* 1986;50:297-317.
- Bosch V, Camejo G. Serum lipoproteinemia in pregnant and lactating rats. *J Lipid Res* 1967;8:138-41.
- Jones CT. Lipid metabolism and mobilization in the guinea pig during pregnancy. *Biochem J* 1976;156:357-65.
- Argiles J, Herrera E. Lipids and lipoproteins in maternal and fetal plasma in the rat. *Biol Neonate* 1981;39:37-44.
- Thomas CR, Lowy C. The interrelationships between circulating maternal esterified and non-esterified fatty acids in pregnant guinea pigs, and their relative contributions to the fetal circulation. *J Dev Physiol* 1987;9:203-14.
- Montes A, Walden CE, Knopp RH, Cheung M, et al. Physiologic and supraphysiologic increases in lipoprotein lipids and apoproteins in late pregnancy and postpartum: possible markers for the diagnosis of prelipemia. *Arteriosclerosis* 1984;4:407-17.
- Ramirez I, Llobera M, Herrera E. Circulating triacylglycerols, lipoproteins, and tissue lipoprotein lipase activities in rat mothers and offspring during the perinatal period: effect of postmaturity. *Metabolism* 1983;32:333-41.
- López-Luna P, Muñoz T, Herrera E. Body fat in pregnant rats at mid- and late-gestation. *Life Sci* 1986;39:1380-93.
- Herrera E, Ayanz A. Calculation of lipolysis and esterification from glycerol metabolism in rat adipose tissue. *J Lipid Res* 1972;13:802-9.
- Chaves JM, Herrera E. In vitro response of glycerol metabolism to insulin and adrenaline in adipose tissue from fed and fasted rats during pregnancy. *Biol Neonate* 1980;38:139-45.
- Llobera M, Montes A, Herrera E. Lipoprotein lipase activity in liver of the rat fetus. *Biochem Biophys Res Comm* 1979;91:272-7.
- Zak B, Cohen J. Automatic analysis of tissue culture proteins with stable Folin reagents. *Clin Chim Acta* 1961;6:665-76.
- Nilsson-Ehle P, Schotz MC. A stable radioactive substrate emulsion for assay of lipoprotein lipase. *J Lipid Res* 1976;17:536-41.
- Corey JE, Zilversmit DB. Validation of a stable emulsion for the assay of lipoprotein lipase activity. *J Lab Clin Med* 1977;89:666-74.
- Lasuncion MA, Testar X, Palacin M, et al. Method for the study of metabolite transfer from rat mother to fetus. *Biol Neonate* 1983;44:85-92.
- Palacin M, Lasuncion MA, Martín del Río R, Herrera E. Placental formation of lactate from transferred L-alanine and its impairment by aminoxyacetate in the late-pregnant rat. *Biochim Biophys Acta* 1985;841:90-6.
- Herrera E, Palacin M, Martín A, Lasuncion MA. Relationship between maternal and fetal fuels and placental glucose transfer in rats with maternal diabetes of varying severity. *Diabetes* 1985;34(suppl 2):42-6.
- Palacin M, Lasuncion MA, Herrera E. Lactate production and absence of gluconeogenesis from placental transferred substrates in fetuses from fed and 48-h starved rats. *Pediatr Res* 1987;22:6-10.
- Freinkel N. Of pregnancy and progeny. *Diabetes* 1980;29:1023-35.
- Herrera E, Knopp R, Freinkel N. Carbohydrate metabolism in pregnancy. VI. Plasma fuels, insulin, liver composition, gluconeogenesis and nitrogen metabolism during late gestation in the fed and fasted rat. *J Clin Invest* 1969;48:2260-72.
- Herrera E, Knopp R, Freinkel N. Urinary excretion of epinephrine and norepinephrine during fasting in late pregnancy in the rat. *Endocrinology* 1969;84:447-50.
- Mampel T, Camprodón R, Solsona J, Junca V, Herrera E. Changes in circulating glycerol, free fatty acids and glucose levels following liver transplant in the pig. *Arch Int Physiol Biochim* 1981;89:195-9.
- Argiles J, Herrera E. Appearance of circulating and tissue ¹⁴C-lipids after oral ¹⁴C-tripalmitate administration in the late pregnant rat. *Metabolism* (in press).
- Childs MT, Tollefson J, Knopp RH, Applebaum-Bowden D. Lipid metabolism in pregnancy. VIII. Effects of dietary fat vs. carbohydrate on lipoprotein and hepatic lipids and tissue triglyceride lipases. *Metabolism* 1981;30:27-35.
- Barter PJ, Gooden JJM, Rajaram OV. Species differences in the activity of a serum triglyceride transferring factor. *Atherosclerosis* 1979;33:165-9.
- Ihm J, Ellsworth JL, Chataing B, Harmony JAK. Plasma protein-facilitated coupled exchange of phosphatidylcholine and cholesteryl ester in the absence of cholesterol esterification. *J Biol Chem* 1982;257:4818-27.

30. Albers JJ, Tollefson JH, Chen ChH, Steinmetz A. Isolation and characterization of human plasma lipid transfer proteins. *Arteriosclerosis* 1984;4:49-58.
31. Herrera E, Gomez-Coronado D, Lasuncion MA. Lipid metabolism in pregnancy. *Biol Neonate* 1987;51:70-7.
32. Borensztajn J, ed. *Lipoprotein lipase*. Chicago: Evener Publishers, 1987.
33. Lasuncion MA, Herrera E. Changes with starvation in the rat of the lipoprotein lipase activity and hydrolysis of triacylglycerols from triacylglycerol-rich lipoprotein in adipose tissue preparations. *Biochem J* 1983;210:639-43.
34. Garfinkel AS, Schotz MC. Lipolytic enzymes and plasma lipoprotein metabolism. *Ann Rev Biochem* 1980;49:667-93.
35. Otway S, Robinson DS. Significance of changes in tissue clearing-factor lipase activity in relation to the lipemia of pregnancy. *Biochem J* 1968;106:677-82.
36. Hamosh M, Cleary TR, Chernick SS, et al. Lipoprotein lipase activity of adipose and mammary tissue and plasma triglyceride in pregnant and lactating rat. *Biochim Biophys Acta* 1970;210:473-82.
37. Lasuncion MA, Herrera E. Effect of pregnancy on the uptake of lipoprotein triglyceride fatty acids by isolated adipocytes in the rat. *Biochem Biophys Res Comm* 1981;98:227-33.
38. Rotherwell JE, Elphick MC. Lipoprotein lipase activity in human and guinea pig placenta. *J Dev Physiol* 1982;4:153-9.
39. Elphick MC, Hull D. Rabbit placental clearing-factor lipase and transfer to the fetus of fatty acids derived from triglycerides injected into the mother. *J Physiol* 1977;273:475-87.
40. Thomas CR, Lowy C, St. Hillaire RJ, Brunzell JD. Studies on the placental hydrolysis and transfer of lipids to the fetal guinea pig. In: Miller RK, Thied HA, eds. *Fetal nutrition, metabolism and immunology: role of the placenta*. New York: Plenum Press, 1984:135-48.
41. Grinberg DR, Ramirez I, Vilaro S, et al. Starvation enhances lipoprotein lipase activity in the liver of the newborn rat. *Biochim Biophys Acta* 1985;833:217-22.
42. Scow RO, Chernick SS, Brinley MS. Hyperlipemia and ketosis in the pregnant rat. *Am J Physiol* 1964;206:796-804.
43. Vilaro S, Reina M, Ramirez I, Llobera M. Intralipid administration induces a lipoprotein lipase-like activity in the livers of starved adult rats. *Biochem J* 1986;236:273-8.
44. Shambaugh GE, Morozak SC, Freinkel N. Fetal fuels. I. Utilization of ketones by isolated tissues at various stages of maturation and maternal nutrition during late gestation. *Metabolism* 1977;26:623-35.
45. Shambaugh GE, Koehler RA, Yokoo H. Fetal fuels. III. Ketone utilization by fetal hepatocyte. *Am J Physiol* 1978;235:E330-7.
46. Desoye G, Schweditsch MO, Pfeiffer KP, et al. Correlation of hormones with lipid and lipoprotein levels during normal pregnancy and postpartum. *J Clin Endocrinol Metabol* 1987;64:704-12.