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# Ultrasound confirmation of ovulation in mares: a normal corpus luteum or a hemorrhagic anovulatory follicle?

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1 Ultrasound confirmation of ovulation in mares: a normal corpus

- 2 luteum or a hemorrhagic anovulatory follicle?

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## **Contents**

The most common pathological anovulatory condition that occurs spontaneously during the breeding season in the mare is the hemorrhagic anovulatory follicle (HAF). A relatively high proportion of mares, soon after ovulation, develop a corpus hemorrhagicum (CH) with a central lacuna. This type of corpora lutea may resemble an HAF, which may complicate the accurate diagnosis of ovulation.. The main objective of this study was to compare the ultrasound data of mares examined frequently with HAFs and CHs to elucidate whether it is possible to distinguish them from each other. A total of 135 ovulating mares were classified according to the morphology of the corpus luteum (CL) in mares with: a solild CL, a CH with small or with large central cavities. Ultrasound characteristics of the development of 11 HAF and 13 CHs with a large central cavity were compared. The preovulatory follicular diameter of ovulatory mares was significantly correlated with the diameter of CH with large central cavities. The percentage of mares with post-ovulatory areas eligible to be mistaken with a CH was less than 25%. Although a predictive diagnosis of an HAF/CH can be made on the basis of several ultrasonographic endpoints, the only parameter that allows a definitive diagnosis is the thickness of the luteal border. This is < 3 mm in HAFs in contrast to > 5 mm in CHs. This only applies when the unidentified structure has non-organized contents.

#### 28 Introduction

In equine practice, the accurate detection and timing of ovulation is important for the following reasons: a) to ensure that ovulation has occurred within the time window

31 considered adequate after pre-ovulation mating; b) to decide the optimum time for 32 breeding with short-lived semen (frozen semen) for post-ovulation AI; c) to determine 33 accurately the embryo age necessary for embryo flushing and recovery; d) to determine 34 the number of ovulations in relation to the number of pre-existent follicles in order to 35 manage any possible twinning properly; and e) to ensure the rupture and collapse of the 36 follicle and so assume proper release of the oocyte in order to differentiate this from 37 pathological anovulatory conditions.

The most common pathological anovulatory condition that occurs spontaneously during the breeding season in the mare is the hemorrhagic anovulatory follicle (HAF) (Ginther et al. 2007; Cuervo-Arango and Newcombe 2010). The main relevance of this condition lies in the failure of the dominant follicle to collapse. It is assumed that the oocyte cannot be released without follicular collapse and fluid evacuation. Therefore it is expected that in mares with HAFs and without normal concurrent ovulation, fertilization is not possible. The mating of 71 mares with solitary HAFs yielded no pregnancy (McCue and Squires 2002). It appears impossible that when using real-time B-mode ultrasonography to distinguish between a preovulatory follicle that will collapse normally and ovulate and another one that will hemorrhage.

For these reasons, it is clinically relevant to be able to diagnose accurately an ovulation and to distinguish it from an HAF. Proper diagnosis of ovulation may be easy to perform if the frequency between examinations is high (e.g. every 12 h) or when the collapse of the follicle results in a solid corpus luteum. A normal ovulation implies the complete collapse of the preovulatory follicle with evacuation of > 90% of fluid (Townson and Ginther 1987). Ultrasonographically, this is visualized as a hypoechoic and solid area in the ovary containing the previous preovulatory follicle during 12 to 15

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h (Newcombe, 1996). This ovulatory area increases subsequently in echogenicity (Townson and Ginther 1989; Newcombe 1996).

However, a relatively high proportion of mares, soon after ovulation, develop a corpus hemorrhagicum (CH) with a central cavity or lacuna of varying dimensions containing fresh blood which later becomes organized (Townson and Ginther 1988; Newcombe 1996). Unfortunately, this type of corpora lutea may resemble an HAF (Ginther et al. 2007), which may complicate the accurate diagnosis of ovulation, especially when mares are not frequently examined for this purpose.

The main objective of this study was to compare the ultrasonographic records of mares examined frequently with HAFs and CHs to elucidate whether it is possible to differentiate the two entities when the mares are examined ultrasonographically at different intervals between examinations. 

#### Materials and methods

#### Animals and ultrasound records

All ultrasound and reproductive data were obtained from mares resident to or visiting a private equine practice in the UK (northern hemisphere) during the breeding seasons of 2009 and 2010. These mares were of various breeds, including Irish Draught, Thoroughbred, Standardbred and Warmbloods. From all data available, ultrasound and reproductive records from a total of 135 mares with ovulatory cycles and 11 mares with HAF cycles were included in the study. Only one cycle per mare was used. The mares with ovulatory cycles had single ovulations and the exact time of ovulation was known within an interval of 8 h. In addition, the ultrasound examinations continued at least every 8 h until two days later (40 to 48 h post-ovulation).

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The ultrasound records included diameters of the preovulatory follicle, the corpus luteum and the HAF at different times relative to ovulation / anovulation; and the appearance of the contents of the HAF's antrum and corpus hemorrhagicum's (CH) central lacuna. The ultrasound examinations were performed by the same operator with a portable scanner (Honda HS-2000V, Honda Electronics Ltd. San Jose, CA, USA) equipped with a 7.5-MHz linear-array transducer.

#### 87 Endpoints analyzed

According to the outcome of the preovulatory follicle, a mare could have an HAF, an ovulation with formation of a solid corpus luteum (solid CL) or an ovulation with formation of a corpus hemorrhagicum (CH). The following end points of these three types of cycle were calculated:

- Follicular diameter: referred to the diameter of the preovulatory follicle 8 h
   before ovulation or before the follicular antrum filled with echoic specks in the
   future HAF (Hour 0). This was measured with the electronic callipers of the
   ultrasound machine by average of two measurements taken at right angles from
   the follicular antrum when the diameter was maximum.
- 97 <u>Ovulation:</u> involved the collapse of the preovulatory follicle with loss of > 90%
  98 of fluid within the following 8 h examination.
- 99 <u>Hemorrhagic anovulatory follicle (HAF)</u>: involved the absence of ovulation,
- 100 entry of echoic particles within the follicle antrum (Hour 0) and gradual increase
- 101 in overall diameter and wall echogenicity. The HAF contents eventually
- 102 organized and did not move freely upon ballottement of the ovary (Fig. 1). The
- 103 overall HAF diameter was calculated from the outer surface of the wall by the
- 104 same technique as for follicular diameter.

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3	105	- Solid corpus luteum and corpus hemorrhagicum: a corpus hemorrhagicum had a
5 6	106	central lacuna $\geq$ 5 mm in diameter within 48 h of ovulation (period during which
7 8	107	the mare was examined). The lacuna contents appeared to be blood. This was
9 10	108	observed initially as many echoic particles floating freely during ballottement of
11 12 13	109	the ovary. Eventually the contents organized forming a network of fibrin strands
14 15	110	(Fig. 2). The diameter of the lacuna increased gradually even after the beginning
16 17	111	of the organization of contents. A solid corpus luteum had no central lacuna. On
18 19	112	some occasions, the solid corpora lutea had a small amount of anechoic fluid (<
20 21 22	113	5 mm; apparently of follicular origin) which usually disappeared by the
23 24	114	following examination. The CH and central lacuna diameters were obtained by
25 26	115	the same technique as for HAFs.
27 28	116	- Organization of contents: The contents of the HAF or central lacuna of the CH
29 30 31	117	were assumed to be blood. They appeared as many echoic particles (too
32 33	118	numerous to count) floating freely during ballottement of the ovary. The term
34 35	119	"organization" or "clotting" of contents referred to the time when at least one
36 37	120	strand of fibrin developed in the central lacuna of the CH or HAF antrum. The
38 39 40	121	strand appeared echoic on ultrasound and was firm and did not move upon
40 41 42	122	ballottement of the ovary.
43 44	123	- <u>Luteal border</u> : corresponded to the luteal tissue of the HAF and CH. In the HAF,
45 46	124	this was measured from the outer surface to the beginning of the HAF antrum. In
47 48 49	125	the CH, it stretched from the outer surface of the CL to the beginning of the
50 51	126	central lacuna. In both structures, the luteal border was more echoic than the
52 53	127	antrum or central cavity, representing luteal tissue.
54 55	128	
56 57 58	129	Experimental design

The percentage of ovulatory mares (n = 135) with a solid CL, CH with central lacuna between 5 and 25 mm and CH with lacuna  $\geq$  26 mm was calculated. The distinction between CHs with small and large lacunae was set at a diameter of 26 mm owing to the minimum diameter of naturally occurring HAFs observed in the present study. In addition, the relationship between the follicular preovulatory diameter and the future CL diameter was determined by correlation analysis. In order to compare ultrasound events between mares with HAFs and CHs for a longer period, all HAF mares (n = 11) and a subset of CH mares with lacunae  $\geq 26$  mm (n = 13) were examined every 8 h until 96 h post-ovulation / anovulation. Furthermore, a final examination was performed 168 h post-ovulation / anovulation to determine the overall diameter after organization and shrinkage of lacuna and HAF contents. **Statistical analyses** All data were tested for normality. Data not normally distributed were ranked for later analysis. The last diameter recorded for the preovulatory follicle (8 h before ovulation) was correlated with the diameter of the CH or solid CL 48 h post-ovulation by Pearson's correlation test. In addition, the preovulatory follicular diameters of the future solid CLs, CHs with small and large lacunae were analyzed statistically by one way ANOVA. The differences in HAF and CH lacuna diameters from Hour 0 to Hour 168 were analyzed by the SAS MIXED procedure with a repeated statement to account for autocorrelation between sequential observations (Version 9.2; SAS Institute, Cary NC, USA). A 2-sample t-test was used to test the difference in several endpoints analysed between HAFs and CHs: maximum diameter, Hour of maximum diameter relative to ovulation /

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155	anovulation, Hour and diameter at which HAF and CH contents organized and the
156	interval from the first evidence of blood to organization of contents.
157	
158	Results
159	
160	Correlation between preovulatory follicles and corpora lutea diameter
161	The 135 ovulating mares developed 43 solid CLs (31.9%) and 92 CHs with a central
162	cavity (68.1%) or lacuna (Table 1). The central lacuna of CHs reached $\geq 26$ mm in
163	diameter in 34.8% of the mares (Table 1). The preovulatory follicular diameter of mares
164	with future CHs with large lacunae ( $\geq 26$ mm) was larger (p = 0.04) than those of mares
165	with future solid CLs and CHs with small lacunae (Table 1). The mean diameter of
166	solid CLs, CHs with small lacunae and CHs with large lacunae at 40 h post-ovulation
167	differed significantly (25.7, 30.5 and 39.1 mm, respectively; p < 0.001). The diameter of
168	CHs with large lacunae at 40 h post-ovulation was positively correlated ( $r = 0.64$ ; $p =$
169	0.001) with the previous preovulatory diameter of its follicles (Fig. 3).
170	
171	General comparisons between endpoints of CHs and HAFs
172	The overall mean diameter of HAFs 0 to 168 h post-ovulation was higher ( $p < 0.001$ )
173	than that of central lacunae from CHs with large cavities (Fig. 4). In both groups, the
174	diameter of HAFs and central lacunae changed over time ( $p < 0.001$ ). There was a
175	significant effect of group by hour interaction on the diameter of HAFs and CHs central
176	lacuna. This effect resulted from a delay in the development of the central lacuna
177	relative to Hour 0 compared with the HAF formation and from an earlier (64 h vs 96 h
178	post-ovulation) beginning of gradual decrease in diameter of the central lacunae
179	compared with that of HAFs (Fig. 4).

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180	The first evidence of ultrasonographic development of central lacunae in CHs relative to
181	Hour 0 varied greatly from 8 to 32 h post-ovulation (Table 2). The proportion of CH
182	central lacunae and HAFs with organized contents (presence of solid fibrin strands)
183	changed over time (Table 2). By Hour 56, all CH central lacunae contents had
184	organized. On the other hand, HAF contents had clotted by Hour 72 (Tables 2 and 3).
185	The luteal border of HAFs was significantly thinner that the border of CHs before
186	organization of contents. Although still significant, this difference became smaller once
187	the contents of both structures had organized (Table 3).
188	The rest of endpoints analysed are shown in Table 3. Overall, there was a great
189	individual variation in the data obtained from both HAF and CH mares. This is
190	indicated by the large range of data points for most parameters analysed. It is worth
191	noting the shorter interval between the first ultrasonographic evidence of blood and the
192	clotting of contents for CHs compared with HAFs (10.4 h and 51.8 h, respectively; $p <$
193	0.001). Furthermore, the mean diameter of HAFs and CH central lacunae at which the
194	contents first developed fibrin strands differed significantly (59.8 mm and 25.3 mm,
195	respectively).
196	
197	Direct comparison between HAFs and CHs at different intervals between
198	examinations
199	- Daily examination: in this scenario the possible youngest and oldest age for HAFs
200	and CHs would be 0 and 24 h, respectively. At this stage, neither HAFs nor CH would
201	have organized contents. In addition, a percentage of future CHs would not have yet a

visible central lacuna (46.1 to 76.9% of ovulating mares; Table 2). In the latter scenario

(collapsed follicle without fluid), there would not be possible misdiagnoses. The two

most accurate criteria that could be used to distinguish between both structures are a)

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205	the thickness of luteal border: in HAFs, the luteal border was always < 3 mm in
206	thickness, while CHs with central lacunae had a luteal border of $\geq 5$ mm in thickness
207	(Table 3); and b) the difference in diameter between the previous preovulatory follicle
208	and the newly developed HAF or CH: the HAFs had a similar or larger diameter than
209	the preovulatory follicle while the CHs was significantly smaller than the preovulatory
210	follicle.
211	- Every other day examinations: the CH and HAF possible ages range between 0 and
212	48 h. At this stage, a percentage of CHs may have developed a central lacuna (46.1 to
213	100%), which would have organized contents in 0 to 92.3% of ovulating mares (Table
214	2). This percentage would increase along with time. As for HAFs, the antrum would
215	have organized contents in 0 to 45.5% of times. An organized HAF would indicate an
216	age of $\geq$ 32 h. When a structure with non-organized fluid is present, the same diagnostic
217	criteria as above apply to differentiate between an HAF and CH. If the unknown
218	structure has organized contents, the best two criteria to distinguish between an HAF
219	and a CH are a) the luteal border thickness, which would be usually, but not always
220	smaller in HAFs than in CHs (Table 3); and b) the difference between the diameters of
221	the preovulatory follicle and the future organized HAF or CH. In the HAF group, this
222	difference is higher (21.5 $\pm$ 3.7 mm; range of 3 to 35 mm; p < 0.001) than that (4.9 $\pm$ 0.9
223	mm; range of $-4$ to 8 mm) in the CH group.
224	- Intervals between examinations longer than two days: at this stage, all
225	ultrasonographic appearances of CHs and HAFs are possible: a) a recently collapsed
226	follicle or a solid CL without fresh blood would undoubtedly indicate follicular rupture
227	and normal ovulation; b) a circular structure with various amounts of fresh blood
228	moving freely upon ballottement of the ovary could indicate either a CH or an HAF. If
229	the luteal border is regular and thinner than 3 mm, it can be ascertained that ovulation

230	did not occur; and finally c) a circular structure with a central network composed of
231	echoic and firm fibrin strands could represent either an organized HAF or CH. In the
232	case of HAFs, the luteal border would be usually thinner than 5 mm, and the overall
233	diameter considerably larger (around 20 mm) than the previous preovulatory follicle.
234	
235	Discussion
236	Occurrence and physiology of fluid-filled luteal glands
237	About two thirds (68.1%) of the ovulating mares included in the present study
238	developed corpora hemorrhagica (CHs) with varying amounts of blood within their
239	central cavities. The proportion of mares with CHs described in the literature agrees
240	with the results of the present study. In two combined studies, Ginther and co-worker
241	found 68.2% (15/22) of mares with CLs with a central clot (Townson and Ginther 1988;
242	Townson and Ginther 1989). In a larger study, Newcombe (1997) reported that 62.4%
243	(118/189) of ovulations developed a fluid-filled central lacunae 42 to 72 h post-
244	ovulation. The reported three studies and the current study also agreed in the great
245	individual variation in terms of timing of central lacuna development relative to
246	ovulation and in the size of the central cavity. This great variation appears to be due to
247	differences in the timing and degree of intraluteal hemorrhage amongst mares.
248	Furthermore, whether a mare develops a solid CL or CH after ovulation seems to occur
249	by chance (Pierson and Ginther 1985). In approximately half of mares with sequential
250	ovulations, a CH formed after one ovulation but not after the other ovulation (Pierson
251	and Ginther 1985). Similarly, in approximately half of the mares that double ovulated,
252	one ovulation developed a CH and the other ovulation a solid CL (Pierson and Ginther
253	1985).

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254	The results of the current study added new information about the relationship between
255	the size of the preovulatory follicle and the final diameter of the CH. This is indicated
256	by the positive and significant correlation between the preovulatory follicular diameter
257	and the CH diameter 40 h post-ovulation. A logical explanation for this correlation
258	could be that a large collapsed follicle would have a larger surface and therefore allow a
259	greater expansion of the newly developing CH as a result of active hemorrhage.
260	The contents of the central cavity remained non-organized for a short period of time
261	(between one and two 8 h-examination intervals). Such a short interval from beginning
262	of hemorrhage to clotting of contents is not surprising since the central cavity of the CH
263	appears to be composed mainly of blood and little, if any, residual follicular fluid. The
264	absence or small amount of follicular fluid, which is rich in a heparin-like substance
265	with anticoagulant properties (Stangroom and Weevers 1962), allows a more rapid
266	fibrinization of the CH contents. The first evidence of clotting was observed as the
267	formation of a solid and echoic strand of fibrin within the CH central cavity. Despite the
268	early organization of the lacuna contents, the size of the central cavity continued to
269	increase to larger diameters. This occurred apparently from further intraluteal
270	hemorrhage, since the ultrasonographic appearance of the CHs with growing lacunae
271	combined the presence of solid strands and echoic particles moving freely upon
272	ballottement of the ovary. The visualisation of these echoic particles is compatible with
273	the presence of fresh blood (Ginther 1992).
274	In contrast to the developmental characteristics of the CH central lacuna, the HAF
275	contents remained in a fluid stage for much longer (32 to 72 h), owing to the higher
276	proportion of follicular fluid in the HAF antrum relative to fresh blood. Furthermore,
277	once the HAF contents organize, the overall diameter reaches a plateau and soon after
278	that, the size of the HAF begins to decrease.

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#### 280 Accuracy of using different ultrasonographic endpoints to distinguish an HAF 281 from a CH

282 If frequent examinations are possible (every 8 to 12 h), the best way to assure that a 283 mare has ovulated is to visualize ultrasonographically the absence of the previously 284 recorded preovulatory follicle. The expected ovulatory site will appear as a hypoechoic 285 area with little or no presence of anechoic follicular fluid (Pierson and Ginther 1985; 286 Newcombe 1996). This indicates the rupture and collapse of the follicle with evacuation 287 of > 90% of follicular fluid and assumes the completion of the process of oocyte 288 release. However, if a longer interval between examination elapses, the presence of a 289 newly formed central cavity with fresh blood within the CH may complicate the 290 accurate diagnosis of follicular collapse. This hypothetical scenario would only occur in 291 less than 23% of the cases. In the remaining proportion of mares (approximately 75%), 292 the accurate diagnosis of ovulation would be possible even if the frequency between 293 two examinations is delayed for 40 to 48 h. This proportion of mares is expected to have 294 a solid CL (with no central blood clot) or a CH with a central lacuna smaller than the 295 possible smallest HAF (< 27 mm in diameter). 296 Unfortunately, the great variation in the maximum diameter of HAFs (42 to 75 mm) 297 overlaps with the maximum CH diameter (35 to 54 mm). And therefore, this 298 overlapping could technically mean a source of error for the practitioner if the criteria 299 for the diagnosis of an HAF/CH are only based on the overall HAF or CH diameter. 300 According to the results of this study there appears to be two ultrasonographic 301 parameters upon which an accurate and definitive diagnosis of an HAF/CH can be 302 based: the luteal border thickness and the difference in diameter between the 303 preovulatory follicle and the future HAF/CH.

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The luteal border of HAF is thinner than that of CHs. The thinner border and therefore the smaller quantity of luteal tissue of HAFs may result from its lack of follicular collapse. The lower concentrations of progesterone in HAF mares involved a reduced vascularisation of developing luteal tissue in unruptured follicles compared with that of corpora lutea which originate from collapsed follicles (Cuervo-Arango et al. 2011). The luteal border thickness is particularly useful in distinguishing between an HAF and CH when their contents remain non-organized. The greater overall HAF diameter compared with the CH's results from the lack of fluid loss in unruptured follicles in addition to the new blood entry that expands the intrafolicular volume of fluid within the HAF. As discussed earlier, the difference between the prerovulatory follicle and the unidentified structure may aid the diagnosis of an HAF, especially at early stages of formation. In conclusion, the preovulatory follicular diameter of ovulatory mares is significantly correlated with the diameter of corpora hemorrhagica with large central cavities ( $\geq 26$ mm in diameter). The percentage of mares with post-ovulatory areas eligible to be mistaken with a CH is less than 25%. Although a predictive diagnosis of an HAF/CH can be made on the basis of several ultrasonographic endpoints, the only parameter that allows a definitive diagnosis is the thickness of the luteal border. This is < 3 mm in HAFs in contrast to > 5 mm in CHs. This diagnostic criterion only applies when the unidentified structure has non-organized contents. If the contents are organized, the luteal border can still be used as a diagnostic criterion though its accuracy is reduced. **Conflict of interest** The authors have no conflict of interest to declare.

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#### 329 Author contributions

- 330 J.R. Newcombe collected the data and J. Cuervo-Arango design the experimental
- 331 protocol and wrote the manuscript up.
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Table 1. Proportion	of mares	with different	luteal	morphol	ogies
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	n	%	POF <sup>A</sup> (mm)	Diam <sup>B</sup> 40 h (mm)	Corr POF- Diam CH
Solid CL	43	31.9	$41.7 \pm 0.8^{a}$	$25.7 \pm 0.6^{a}$	0.37
Lac 5-25	60	44.4	$42.0 \pm 0.7^{a}$	$30.5 \pm 0.6^{b}$	0.39
$Lac \ge 26$	32	23.7	$44.7 \pm 1.1^{b}$	$39.1 \pm 1.1^{\circ}$	0.64
P-value			0.04	0.001	

A: Preovulatory follicular diameter (POF) 8 h before ovulation of mares with solid corpora lutea (CL), with corpora hemorrhagica (CH) and central lacuna of 5 to 25 mm and  $\geq$  26 mm in diameter. B: Diameter of CLs and CHs (outer ring) 40 to 48 h post-ovulation.

C: Pearson's correlation coefficient (r) between the POF and the CL/CH diameter 40 to 48 h post-ovulation.

Table 2. Ultrasonographic events of HAF and CH lacuna contents

Hours post-	% of CH with	Diameter of	% of CH with	% of HAF with
ovulation/anovulation	visible lacuna	lacuna (mm)	clotted lacuna	clotted contents
(h)				
0	46.1% (6/13)	3.5 ±0.5	0.0% (0/6)	0.0% (0/11)
8	30.1% (4/13)	13.2±1.2	0.0% (0/4)	0.0% (0/11)
16	53.8% (7/13)	17.6±2.3	2.3% (1/13)	0.0% (0/11)
24	76.9% (10/13)	21.3±3.6	23.1% (3/13)	0.0% (0/11)
32	100% (13/13)	24.8±3.4	61.5% (8/13)	27.3% (3/11)
40	100% (13/13)	30.1±3.0	84.6% (11/13)	36.4% (4/11)
48	100% (13/13)	34.1±2.1	92.3% (12/13)	45.4% (5/11)
56	100% (13/13)	36.8±1.7	100% (13/13)	63.6% (7/11)
64	100% (13/13)	36.8±2.1	100% (13/13)	81.8% (9/11)
72	100% (13/13)	35.3±2.1	100% (13/13)	100% (11/11)

Table 3. Ultrasonographic endpoints of HAF and CH with central lacunae  $\geq$  26 mm in diameter

	n	Max <sup>A</sup> diam (mm)	Luteal border <sup>B</sup> (mm)	Luteal border <sup>C</sup> (mm)	Hour of <sup>D</sup> max diam (h)	Diam at <sup>E</sup> clotting (mm)	Hour of <sup>F</sup> clotting (h)	Interval <sup>G</sup> hemorrhage- clotting (h)
HAF	11	61.0±3.4	2.1±0.1	4.3±0.3	74.2±4.5	59.8±3.3	51.8±4.8	51.8±4.8
(range)		42-75	1.5-2.5	3.0-5.5	60-96	40-73	32-72	32-72
CH	13	47.5±1.5	7.9±1.9	6.5±1.3	61.8±5.5	25.3±2.1	39.6±3.7	10.4±1.2
(range)		35-54	5.0-11.0	4.5-9.0	40-96	17-28	16-56	8-16
P-value		0.003	0.001	0.03	NS	0.001	0.01	0.001

A: Overall maximum diameter (outer ring) of hemorrhagic anovulatory follicles (HAF) and corpus hemorrhagicum (CH) with a central lacuna  $\geq 26$  mm in diameter.

B: Luteal border of HAF and CH with non-organized (fresh blood moving freely upon ballottement of the ovary) contents at its maximum diameter.

C: Luteal border of HAF and CH with organized contents at its maximum diameter.

D: Hour relative to Hour 0 (hour of ovulation / anovulation) at which the HAF and CH reached the maximum diameter.

E: Diameter of HAF and CH central lacuna at which the contents organized

F: Hour relative to Hour 0 at which the contents of HAF and CH central lacuna organized

G: Interval in hours between from the first evidence of fresh blood in HAF and CH to the moment of organization of contents.

Difference <sup>C</sup>

	n	POF <sup>A</sup> (mm)	Diam flu contents (
HAF	11	$38.5 \pm 1.8$	39.2±2
(range)		(26/49)	(27/49
CH	13	$42.5 \pm 1.5$	19.9 ±
(range)		(34/51)	(10/28
D volue		0.03	0.001

reovulatory follicles and CH /HAF

s (mm) (mm) ±2.2  $0.7 \pm 1.8$ (49) (-11/8) $-22.5 \pm 1.2$ ± 1.4 (-30/-15)(28) 0.001

e ovulation or the formation of hemorrhagic anovulatory

maximum diameter at which its contents remain nonorganized.

C: Difference between the diameter of A and B.

from CHs wn... All CH data are from CHs with lacunae > 26 mm in diameter.





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Hours after ovulation / anovulation

Fig. 4

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Fig. 1. Representative B-mode ultrasonograms of a mare that failed to ovulate and developed a hemorrhagic anovulatory follicle (HAF).

Fig. 2. Representative B-mode ultrasonograms of a mare with an ovulation and subsequent corpus hemorrhagicum development with a growing central lacuna 0 (ovulation) to 168 h post-ovulation. Note in pictures "0 h" and "8 h" the presence of residual follicular fluid mostly anechoic. In picture "16 h" the central lacuna is composed of many echoic particles moving freely during ballottement of the ovary. This appearance is compatible with fresh blood. In picture "24 h", the central lacuna contents begin to organize: a solid and echoic fibrin strand can be observed across the center of the lacuna.

Fig. 3. Scatter plot diameters of preovulatory follicles and diameters of corpora hemorrhagica (outer ring) with central lacunae  $\geq$  26 mm 40 to 48 h post-ovulation (n = 32). The Pearson's correlation coefficient was significant (p < 0.001) and indicated a positive correlation (r = 0.64) between the POF and the CH diameters two days later.

Fig. 4. Mean diameter ± SEM of preovulatory follicles, hemorrhagic anovulatory follciles (n = 11), corpora hemorrhagica (n = 13) and central lacunae (n = 13) – 24 to 168 h postovulation/anovulation. Only mares with CHs and central lacunae  $\geq$  26 mm in diameter were included. The effect of group (HAF vs. central lacuna of CHs), effect of Hour and effect of group by hour interaction on the diameter of HAF and central lacuna was significant (G: p < 0.001; H: p < 0.001; G\*H: p < 0.05, respectively).

R.