



## Review Article

## Biochemical and Hematological Indexes of Liver Dysfunction in Horses

Katuska Satué<sup>a,\*</sup>, Esterina Fazio<sup>b</sup>, Pietro Medica<sup>b</sup>, Laura Miguel<sup>a</sup>, Juan Carlos Gardón<sup>c</sup><sup>a</sup> Department of Animal Medicine and Surgery, Faculty of Veterinary Medicine, CEU-Cardenal Herrera University, Valencia, Spain<sup>b</sup> Department of Veterinary Sciences, Veterinary Physiology Unit, Polo Universitario Annunziata, Messina, Italy<sup>c</sup> Department of Animal Medicine and Surgery, Faculty of Veterinary and Experimental Sciences, Catholic University of Valencia-San Vicente Mártir, Valencia, Spain

## ARTICLE INFO

## Article history:

Received 27 February 2023

Received in revised form 17 March 2023

Accepted 17 March 2023

Available online 22 March 2023

## Keywords:

Biochemical

Hematological

Interpretation

Hepatic function

Horse

## ABSTRACT

In the present review, the authors, based on the multiple functions performed by the liver, analyze the multiple biochemical and hematological changes as an expression of altered liver function in the horse. The liver performs important metabolic functions related to the synthesis, degradation, and excretion of various substances. Modification of these functions can be evaluated and diagnosed by determining serum concentrations of several serum analytes, including enzymes and other endogenous substances. Hepatocellular enzymes, such as sorbitol dehydrogenase-SDH and glutamate dehydrogenase-GLDH, are released following hepatocellular necrosis. Hepatobiliary enzymes, such as  $\gamma$ -glutamyl transferase-GGT, increase in response to necrosis, cholestasis, and other alterations in bile conducts. Serum concentrations of mainly endogenous and exogenous substances that the liver should synthesize or eliminate, such as proteins (albumin and globulins), bile acids, urea, glucose, total and direct bilirubin, and coagulation factors, and fibrinogen should be included in the liver function test profile. The interpretation of laboratory tests of liver function will allow the diagnosis of functional loss of the organ. Some of the analytes considered provide information on the prognosis of liver disease. This review will provide an accurate and objective interpretation of the common biochemical and hematological tests in use in the diagnosis of equine hepatic disease patients, aiding still further the veterinary activity on the applied equine clinical cases.

© 2023 The Authors. Published by Elsevier Inc.

This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)

## 1. Introduction

The functional liver failure's syndrome covers a board spectrum of clinical, biochemical, and neurophysiological changes. Hepatic failure or insufficiency refers to the liver inability to perform its physiological functions, characterized by deterioration in the synthesis, regulation, and detoxification function [1]. Because the liver is involved in such a wide variety of physiologic functions, any pathologic process may hinder one or several activities without impeding others. What is more, the high reserve capacity of the liver induces it capable of remaining functional albeit large sum of tissue damage in the horse [2]. Despite the ability to regenerate, death, or necrosis of liver cells causes them to be re-

placed with fibrous tissue. The correct and incontrovertible etiology of chronic liver disease in equine adults, by considering analysis of clinical, ultrasonographic, serum biochemical, hematological, and immunoreactivity data, is rarely identified based in the biochemistry; however, some causes can include cholangiohepatitis, aflatoxicosis, and pyrrolizidine alkaloid toxicity (*Senecio* spp.), alsike clover, iron toxicity [3], and viral hepatitis as equine hepatitis virus (EqHV) [4] and equine parvovirus hepatitis (EqPV-H) in adult horses [5,6], among others [1,7]. Replacement with fibrous tissue is a serious problem and carries out a poor prognostic evaluation since the hepatic function is compromised. However, the prognosis of equine hepatic insufficiency depends upon the type and related severity of the underlying disease. Unfortunately, hepatic diseases and liver failure are difficult to diagnose in the early stages, until more than 60% to 80% of the liver is non-functioning [19].

In horses, the most common clinical signs of hepatic insufficiency are represented by, anorexia, weight loss, icterus, fever, hepatic encephalopathy, and depression. On the other hand, the less commonly reported clinical signs include diarrhea, colic, hepatogenic photosensitization, hemorrhagic diathesis, polycythemia, and

Conflict of interest statement: The authors declare no conflicts of interest.

Animal welfare/ethical statement: Ethical approval is not necessary for this kind of study.

\* Corresponding author at: Katuska Satué, Department of Animal Medicine and Surgery, Faculty of Veterinary Medicine, CEU-Cardenal Herrera University, Valencia 46115, Spain.

E-mail address: [ksatue@uchceu.es](mailto:ksatue@uchceu.es) (K. Satué).

bilateral laryngeal paralysis. In addition, polydipsia, hemolysis, dependent abdominal edema, ascites, tenesmus, and endotoxic shock are rarely possible to observe. The appearance of specific clinical signs of hepatic diseases often reflects the typology of physiological or altered hepatic functions [8,9,19]. Since the functions of the liver are numerous, its diseases and/or failure can affect the fat, protein, and carbohydrate metabolism, the excretion of bile, breakdown and excretion of compounds, toxins' filtration, aged erythrocytes and organisms, storage of vitamins (A, D, and B12) and minerals (copper and iron); among others, changes in coagulation factors, fibrinogen, ammonia, urea (BUN), total bile acids (TBA), bilirubin (BIL), albumin (ALB) and globulins (GLOBs) could indicate although not consistently in all liver dysfunction patients (Mair et al., 2017). These liver function test only become abnormal when approximately 60 - 70% of liver function is lost [19].

Blood tests are generally very helpful in clinical diagnosis of equine hepatic diseases, but they have a limited prognostic value. In the analysis of the liver profile, some biochemical indices are helpful for prognoses, such as significantly elevated levels of GLOBs (x 13 risk of death at 6 months) and TBA (x 10 risk of death at 6 months) and significantly reduced levels of ALB (x 10 risk of death at 6 months) [10].

Bile acids (BA) are a group of amphipathic steroids synthesized by hepatocytes from cholesterol and excreted into bile. They function to emulsify fat in intestine and facilitate nutrient absorption and are highly conserved via enterohepatic circulation as outlined below [11]. BA concentrations above the reference interval indicate a poor prognosis in UK horses with liver disease [10,12,13]. An experimental study, carried out in 9 horses, evaluated the toxicity of the pyrrolizidine alkaloid, resulting in an increase of BA above 50  $\mu\text{mol/L}$  in 8 of them, which were euthanized for the severity of the disease or died spontaneously; the only surviving horse showed a BA concentration of 33  $\mu\text{mol/L}$  [16]. Delvescovo et al. [14], showed that the survival rate in horses with BA >30  $\mu\text{mol/L}$  was 55%, which is in line with the previously reported 52% survival for cases with BA concentrations of >20  $\mu\text{mol/L}$  [1], confirming the guarded prognostic indicators for all these cases. Moreover, the degree of BA elevation above 30  $\mu\text{mol/L}$  concentrations did not worsen the prognosis for survival up to 6 months after hospital release.

Survival prognosis improves when liver damage is reversible and, in these cases, a single measurement of BA might not provide enough information about the potential for recovery and survival. The prognostic value of BA relies on the fact that the detected loss of hepatic function is permanent or progressive or both. It is conceivable that reversible changes, such as inflammatory infiltration or reversible hepatocyte damage, could temporarily interfere with hepatic function, leading to an increase in BA. In contrast, in cases of serum hepatitis, cholangiohepatitis, neoplasia, Clostridium piliformis infection, and pyrrolizidine alkaloid toxicity are all associated with severe or non-reversible liver damage, a single BA value might be more predictive of survival [14]. In many of these cases, loss of hepatic function is permanent or progressive, or both, and BA are likely to be indicative of extent of the irreparable damage and therefore also of the prognosis. In these cases, a close association between BA, presence and severity of hepatic damage and a negative outcome would be expected [12]. However, BA returned to normal levels in horses that survived liver necrosis from mild to moderate liver disease of unknown etiology, and in these cases, BA might be more indicative of a temporary and potentially reversible compromise of hepatic function corresponding to improved liver function as regeneration occurred [15]. This assumption is supported by the low specificity of BA >17 and >20  $\mu\text{mol/L}$  for nonsurvival and the correlation between BA and potentially reversible histological findings such as inflammation and hemosiderin accumulation in Kupffer cells suggesting interference with hepatic function [12].

**Table 1**

Common findings of hepatobiliary enzymes associated with liver functional abnormalities [8,13,6,4].

Specific Enzymatic Indicators of Liver Disease		
Enzyme	Dynamic	Range (UI/L)
SDH	> 8	2-8
GGT	> 20	5-20
GLDH	> 10	2-10
Arginase	> 4.2	0-4.2

In these latter studies, the predictive value of survival based on pyrrolizidine alkaloid toxicity could not be established for the small number of horses used in the study (only 3 cases). However, BA concentrations exceeded 50  $\mu\text{mol/L}$  and these animals died or were euthanized for the severity of the disease. This supports the hypothesis observed by [16] that high BA concentrations (>50  $\mu\text{mol/L}$ ) are associated with the nonsurvival of this disease. In contrast, in cholangiohepatitis and hepatic lipidosis, the survival rate was 79% (15/19 survivors) and 87% (7/8) despite the marked elevations in BA concentrations present in some cases. These findings suggest that the primary disease and the potential for reversible liver damage are important in evaluating the prognostic indicator of elevated BA concentrations. The relationship between prognosis and direct bilirubin (DBIL) has only been reported by Dunkel et al. [12], although in this study only DBIL was measured in 2 horses that did not survive, resulting in low statistical power; and indirect bilirubin (IBIL) was not assessed due to confounding effects of hemolysis and anorexia. Based on the results recorded by Delvescovo et al. [14] the magnitude of the elevation of DBIL could represent a better prognostic indicator than BA concentrations in horses from the eastern United States with hepatic disease and BA >30  $\mu\text{mol/L}$ , furthermore, the DBIL concentration was positively correlated with non-survival.

Therefore, an approach to interpreting abnormal liver function through changes in main biochemical and hematological markers in order to aid subsequent diagnosis was considered in this review.

## 2. Biochemical Profile of the Equine Hepatic Function

Since before alterations in some laboratory parameters are observed, the liver injury must be massive and since pathological processes can differentially alter liver functions, the laboratory diagnosis of liver failure can be a challenge. Increased serum activities of several intracellular enzymes have been reported to be useful in establishing the diagnosis and prognosis of hepatopathies in the horse [5,17]. Hepatocellular enzymes located in the cytoplasm, like sorbitol dehydrogenase-SDH and arginase or the mitochondria, as soon as glutamate dehydrogenase-GLDH, are released following hepatocellular necrosis. Serum activity of these enzymes depends on the number of hepatocytes injured, the severity of the disorders, and the half-life of the enzyme activities involved in both acute and chronic disease. In fact, since the half-life is very short for the enzymes LDH-5 (<24 hours), SDH (<12 hours), and GLDH (12-14 hours), the increase due to hepatocellular injury is followed by a rapid decrease. Thus, repeated measurements of these enzymes may be useful in determining the progression of acute hepatocellular injury, but less appropriate for the diagnosis of chronic liver injury [8,18]. On the other hand,  $\gamma$ -glutamyl transferase-GGT is mainly released from the biliary epithelium and in cases of inflammation, biliary hyperplasia, or destruction of the epithelium of the bile canaliculi and secondary to the inflammation of cholestatic hepatocytes, it continues to rise for a few days, although liver damage is not present [19]. However, in horses, increased GGT activity may also be associated with hepatocellular damage and liver necrosis [5,17-19] (Table 1).

The identification of increased hepatic-derived enzymes is used to diagnose liver damage, but the magnitude of the increases is not sufficiently reliable to predict prognosis [18]. Prognosis is best assessed by function test abnormalities, in conjunction with the identification of the etiology and the presence or absence of hepatic encephalopathy [1].

Other enzymes located in hepatocytes (aspartate aminotransferase-AST and lactate dehydrogenase isoenzyme 5 (LDH-5) and in the epithelium of the bile ducts (alkaline phosphatase-ALP), although they are often included in hepatobiliary profiles in horses, are not specific for hepatobiliary damage since they can be found in other locations. In fact, AST is also expressed in striated muscle, so other liver-specific enzymes (SDH or GLDH) or muscle-specific enzymes such as creatine kinase (CK) must be used to differentiate between hepatocellular or muscle damage. While the marked increase in both AST and SDH suggests acute or active hepatocellular injury, the increase in AST and CK is indicative of muscle damage. Likewise, ALP is also expressed in bone, kidney, intestine, and placenta, and LDH-5 in kidneys, muscle, myocardium, and red blood cells [1]. In copper-associated liver cirrhosis elevations of ALP, AST, GGT, LDH, and BA and prolongation of Prothrombin Time (PT) were identified. Although the cause of hepatic copper accumulation was not identified, Ankringa et al. [20] concluded that excess copper in the liver should be considered in the differential diagnosis of hepatic cirrhosis in the horse. However, in chronic hepatic disease with severe liver fibrosis and despite the reduction of its functionality, the serum activities of hepatic enzymes might remain in the physiological range [2].

Serum concentrations of mainly endogenous and exogenous substances that the liver should synthesize or eliminate (ALB, GLOBs, BA, BUN, GLU, TBIL and DBIL, fibrinogen (FIB) and coagulation factors should be included in the liver function test profile [21,18,22]. Since hepatic tests of functionality become abnormal only after 70% or more of its function is lost [22], biochemical indicators can help narrow the differential diagnosis for hepatic failure and, when evaluated over time, can help predict prognosis. Specific parameters, indicators of hepatic disease, include BA, DBIL, ammonia concentrations, and the ratio between branched amino acids: aromatic amino acids (BCAA/AAA ratio). Nonspecific parameters indicators of liver disease include TBIL, IBIL, BUN, GLOBs, ALB, GLU, prothrombin time (PT) and activated Partial Thromboplastin Time (aPTT), triglycerides (TGs), and FIB.

### 2.1. Bile Acids

Bile acids (BA) are present in high concentrations in the portal circulation, are extracted efficiently by the liver, and are normally transported away from the hepatic tissue through the biliary circulation. The enterohepatic circulation normally removes more than 90% of BA concentrations [23]. It is estimated to cycle approximately 40 times a day in ponies [83]. The determination of the total BA (TBA) concentration is considered a highly sensitive assay in cases of hepatic dysfunction, and its increase is an excellent indicator of liver failure but is not specific to the type of liver disease in horses [15]. Normal TBA concentrations in healthy adult horses between 5 and 28  $\mu\text{mol/L}$  have been reported with neither diurnal variations and postprandial increase, nor time-of-day variations [24]. TBA concentrations  $>20 \mu\text{mol/L}$  have high sensitivity and positive predictive value for hepatic diseases in horses [1,13]. Indeed, horses with chronic liver disease and persistently increased TBA concentrations  $>20 \mu\text{mol/L}$  have a guarded to poor prognosis [12,1]. Survival rates for horses with TBA  $>20 \mu\text{mol/L}$  [1] and  $>30 \mu\text{mol/L}$  [14] have been reported to range from 52 and 55%, respectively, which indicates the reserved prognosis in these cases. Milder increases (up to  $20 \mu\text{mol/L}$ ) may occur in a few horses

without hepatic disease that result anorexic for 2 or more days (20–30  $\text{mmol/L}$ ) [25]. However, TBA concentrations should not be used as a predictor of prognosis in horses with acute hepatic disease.

An experimental study, carried out in 9 horses, evaluated the toxicity of the pyrrolizidine alkaloid, resulting in an increase of BA above  $50 \mu\text{mol/L}$  in eight of them, which were euthanized for the severity of the disease or died spontaneously; the only surviving horse showed a BA concentration of  $33 \mu\text{mol/L}$  [16]. Delvescovo et al. [14], showed that the survival rate in horses with BA  $>30 \mu\text{mol/L}$  was 55%, which is in line with the previously reported 52% survival for cases with BA concentrations of  $>20 \mu\text{mol/L}$  [1], confirming the guarded prognostic indicators for all these cases. Moreover, the degree of BA elevation above  $30 \mu\text{mol/L}$  concentrations did not worsen the prognosis for survival up to 6 months after hospital release.

In foals under 1 week of age, the interpretation of TBA concentrations is difficult. Compared with those of healthy adult horses, TBA concentrations in healthy foals are considerably higher during the first 6 weeks of life. In clinically ill neonatal foals, it is particularly important to have healthy controls of the same age or reference values concerning age [83].

### 2.2. Total, Direct, and Indirect Bilirubin

In normal horses, the TBIL concentration varies between 0.2 and 5.0  $\text{mg/dL}$ , with DBIL in the range of 0 to 0.4  $\text{mg/dL}$  [14]. Failure of the liver to absorb, conjugate, and/or excrete BIL can lead to increases in IBIL and/or DBIL. However, both anaemia and hemolysis are causes of unconjugated hyperbilirubinemia in healthy horses [15]. The latter condition may be caused by a congenital deficiency of glucuronyl transferase and affected horses may maintain persistent TBIL concentrations of 9  $\text{mg/dL}$  or more [26].

Most cases of equine hepatic disease present moderate increases in TBIL concentration (typically 50–150  $\text{mmol/L}$ ) in which the IBIL often greatly exceeds the DBIL fraction. However, the increased concentrations of IBIL, which occurs in equine hepatic diseases, are associated with acute hepatocellular but do not necessarily indicate the presence of hepatic failure. Similarly, TBIL values may be within physiological ranges in animals with severe chronic disorders of the liver [27].

According to Barton and LeRoy [83] and DeNotta and Divers [28] when serum DBIL concentration is  $>25\%$  of the TBIL value, the hepatocellular disease should be considered in the differential diagnosis, and when  $>30\%$  of the total value, cholestasis should be considered as well. DBIL concentration is water soluble and detectable in horse's urine (bilirubinuria) only if blood concentrations rise high enough to exceed the renal threshold; therefore, when urine tests are positive for the presence of BIL, cholestatic disease should be suspected [14].

In case of liver damage in horses, most of the BIL is unconjugated and the DBIL/TBIL ratio is usually  $<0.3$ . In horses with acute liver failure, the IBIL is the fraction that increases as the IBIL rarely exceeds 25%–35% of the TBIL.

### 2.3. Ammonia

Physiological values for ammonia in the range of 13 to 108  $\mu\text{g/dl}$  in healthy horses have been reported. Blood ammonia concentrations can range from slightly above normal to more than 1000  $\mu\text{mol/L}$ , and higher concentrations at admission were associated with nonsurvival in one retrospective report including 36 cases [29].

Hyperammonemia contributes to hepatic encephalopathy in equine patients with liver failure; however, hyperammonemia can

also be caused by gastrointestinal (GI) disorders, poisoning, or in-born metabolic defects. However, patients with hepatic failure have other metabolic complications that do not occur in patients with other causes of hyperammonemia. Hyperammonemia results when ammonia from portal circulation is not metabolized to BUN within the liver and enters the systemic circulation. Raised plasma ammonia affects gluconeogenesis, causing the utilization of branched-chain amino acids. The resultant decrease in the ratio of branched chain to aromatic amino acids (BCAAs to AAAs ratio) affects neurotransmission and may give rise to hepatic encephalopathy or, very rarely, peripheral neuropathy which can lead to unusual complications such as gastric impaction (and colic) or laryngeal paralysis [30].

Although circulating ammonia concentrations can be increased in equine patients with hepatic failure, they do not correlate well with the severity of hepatic encephalopathy, except in portocaval shunts [31]. Hyperammonemia with clinical signs of encephalopathy is occasionally also recorded in horses with acute GI disease, without evidence of concurrent hepatic diseases [29,32,33]. In such cases, hyperammonemia is caused by the increased production of ammonia in the GI tract in combination with increased gut permeability that facilitates ammonia absorption in the horse [34].

Increased blood TBA and ammonia concentrations, with hepatic enzymes' activity usually normal, and signs of hepatic encephalopathy without liver failure have been re-reported in Morgan horses at weaning; this profile, related to an inherited defect in a trans-porter protein at the mitochondrial level that, causes deficiency in ornithine, necessary for urea synthesis, has been identified. This process is characterized by a syndrome of hyperammonemia, hyperornithinemia, and normocitrullinuria [35].

#### 2.4. Branched and Aromatic Amino Acids Ratio

Serum concentrations of branched-chain amino acids (BCAA), leucine, isoleucine, and valine are decreased, while the concentrations of the aromatic amino acids (AAAs) phenylalanine, tyrosine, and tryptophan are increased in patients with advanced liver diseases, resulting in a low of BCAAs/AAAs ratio of plasma amino acids in human cirrhotic patients [36]. A low ratio of branched and aromatic amino acids has been associated with hepatic encephalopathy; hence, a decrease below 2.5 in this ratio has been associated with an increased risk of developing signs of hepatic encephalopathy in horses [37,38]. The imbalance of amino acids tends to become more marked with the progression of hepatic diseases, and aminograms are useful for assessing the prognosis of human cirrhotic patients with or without hepatocellular carcinoma, as have been showed in humans [39].

#### 2.5. Urea and Creatinine

Because the liver is primarily responsible for removing ammonia from the circulation and converting it to BUN for renal excretion, increases in blood ammonia concentration or a decrease in BUN concentration (< 9 mg/dL) may be indicative of chronic hepatocellular disease in the horse [40,83]. Decreased plasma BUN is associated with severe hepatopathies, therefore in these conditions, the measurement of serum BUN concentration has a prognostic value and might be served as a useful screening test for the detection of hepatic insufficiency in these horses [2]. Creatinine is also sometimes low in the liver disease cases for unknown reasons but perhaps due to washout associated with polydipsia. Low concentrations of creatinine in human cirrhotic patients have been related to the presence of protein-calorie malnutrition, reduced muscle mass, altered creatinine production in the liver, and increased excretion of creatinine in the renal tubules of human patients [41].

Although in horses, decreased liver function, advanced age, and reduced muscle mass can maintain creatinine concentration within the physiological range, animals hospitalized for acute liver disease may show prerenal azotemia secondary to dehydration and hypovolaemia, such as colic or diarrhoea [42].

#### 2.6. Albumin and Globulins

Since ALB has a long plasma half-life (19–20 days), hypoalbuminemia in horses is rare, and it is more common in horses with chronic (18%) than acute (6%) hepatic disorders. Indeed, a decreased functional hepatic parenchyma most likely explains the low ALB concentration in patients with chronic hepatic disease. Since concentrations < 2.0 g/dL are not common even in severe equine liver disease [1], hypoalbuminemia is neither a sensitive nor a specific test for hepatic failure in the horse. ALB concentration in foals is significantly higher in the neonatal foals, on the day of the birth (less than 12 hours old), so it is important to use age-based references to assess liver profile during this early period [43,44].

In contrast, GLOBs are increased in 48% of horses with liver failure. Hyperglobulinemia is probably produced because of systemic immunostimulation by gut-derived antigenic material following the loss of Kupffer cells in the liver [45]. Hyperglobulinemia (polyclonal gammopathy or increased  $\beta$ -globulins) can develop in horses with severe acute or chronic liver disease. Although the total protein concentration may be normal, the albumin/globulin ratio is usually decreased [1,46]. Hyperglobulinemia, in association with other clinicopathologic indicators of liver disease, is an important indicator of marked liver injury, and the magnitude of the increase in serum globulin concentration has prognostic relevance. Serum globulin concentrations greater than 45 g/L are worrisome and values up to 60 to 70 g/L justify a guarded prognosis [1]. However, it must be kept in mind that the increase in GLOBs must be interpreted in conjunction with other indicators of liver disease, such as liver enzymes, to support the diagnosis of liver disease/insufficiency.

However, hypergammaglobulinemia has also been associated with monoclonal gammopathy in B-cell lymphomas in horses. In fact, Badial et al. [47] on histopathological examination of tissue samples collected during necropsy in three horses revealed neoplastic lymphocytes in the lymph nodes and perivascular infiltrates in the intestinal tract, kidney, heart, and liver. These three animals were diagnosed with B-cell lymphoma characterized by small to intermediate lymphocytes with concurrent tissue and blood involvement and monoclonal gammopathy of IgG isotypes.

Commonly, impaired liver function reduces the capacity for protein synthesis, so that the ALB and  $\alpha$ - and  $\beta$ -GLOB fractions decrease. However, total GLOBs may increase due to increased  $\alpha$ -GLOB production as part of the acute phase response,  $\beta$ -GLOB production under inflammatory conditions, liver disease, or parasitism; and  $\gamma$ -GLOB by chronic antigenic stimulation associated with infection. While polyclonal gammopathy is common in chronic bacterial infections or autoimmune pathologies or liver cirrhosis [48], some forms of neoplasia (lymphoma or myeloma) can cause monoclonal gammopathy in the region of  $\beta$ - or  $\gamma$ -GLOBs ([49–51]; Gianetto et al., 2022).

Common variable immunodeficiency (CVID) is a rare condition in adult horses characterized by hypogammaglobulinemia and increased susceptibility to parasitic and bacterial infections, including recurrent respiratory diseases, septicemia, hepatitis, and meningitis. B. burgdorferi has been speculated to be the cause of hepatitis in other cases of equine neuroborreliosis [52]. Indeed, Pecoraro et al. [53] identified one case of neuroborreliosis associated to Borrelia burgdorferi in a CVID horse related with hepatitis.

**Table 2**

Common findings of biochemical parameters and coagulation times associated with liver functional abnormalities ([13]; Dunkell et al., 2011; [31,57]).

Specific Biochemical Indicators of Liver Disease		
Parameter	Dynamic	Range
TBA	> 15	< 15 $\mu\text{mol/L}$
DBIL	> 0.4	0.1–0.4 mg/dL
DBIL/TBIL	> 0.2	< 0.2
Ammonia	> 108	13–108 $\mu\text{mol/L}$
BCAA/AAA ratio	< 3.5	3.5–4.5
Bilirubinuria	Positive	Negative
Nonspecific Biochemical Indicators of Liver Disease		
TBIL	> 2.1	0.5–2.1 mg/dL
IBIL	> 2.0	0.3–2.0 mg/dL
BUN	< 10	10–29 mg/dL
GLOBs	> 3.8	2.3–3.8 g/dL
ALB	< 2.9	2.9–3.6 g/dL
GLU	< 71	71–122 mg/dL
PT	> 14	12–14 sec
APTT	> 35	24–35 sec
TG	> 65	14–65 g/dL
FIB	> 400 (inflammation) < 200 (liver chronic disease)	200–400 mg/dL

### 2.7. Glucose

Although changes in blood GLU concentrations are rarely observed in horses with chronic hepatic disease, hypoglycemia (GLU < 60 mg/dL) can occur in massive acute liver failures of horses [54,55], with liver necrosis, lipidosis, neoplasia, and cirrhosis, being more frequent in foals with hepatic failure than in adult horses. It is likely that as anorexia progresses in liver failure, glycogen stores are depleted, and thus gluconeogenesis and subsequent glycolysis occur from increased glucagon secretion [13]. Hypoglycemia, associated with chronic liver disease in horses, is indicative of a guarded prognosis [1].

### 2.8. Clotting Parameters

Decreased synthesis of most pro-coagulant, anticoagulant, and fibrinolytic proteins, as well as reduced platelet number and function, are associated with hepatic failure in the horse. Thus, the evaluation of haemostatic function requires the determination of platelet count, prothrombin time (PT), activated partial thromboplastin time (APTT) and fibrinogen concentration, and fibrin degradation products (FDPs). The reduction in the synthesis of factors II, V, VII, IX, X, XI, and XII is represented at the laboratory level by the prolongation of the PT and APTT. Because factor VII has the shortest half-life and is dependent on vitamin K, abnormalities are seen primarily in PT. In general, a 50% to 70% decrease in the concentration of these factors is necessary for changes to be detectable. Peek and Divers [56] concluded that coagulation abnormalities are not always detectable in all horses with hepatic disease, including obstructive biliary disease and insufficiency. This could be contradictory since it is well known that BA and vitamin K are necessary for the synthesis of coagulation factors II, VII, IX, and X and inhibitors of proteins C and S [56]. Natural anticoagulants such as antithrombin III (AT III) and proteins C and S are also produced in the liver. Decreased hepatic clearance of plasminogen activators, such as tissue plasminogen activator, can result in fibrinolysis [45]. In a population of 65 horses, Johns and Sweeney [57] identified at least one abnormality in a routine coagulation profile (platelet count, PT and APTT times, AT III activity, fibrin degradation products (FDPs), and FIB concentration) in 58% of horses. Although coagulation abnormalities occur in horses with liver disease, these investigators observed no clinical evidence of a bleeding diathesis. Surprisingly, in those horses with fulminant liver disease, platelet counts remained within reference intervals.

Although bleeding is not the most common clinical manifestation in these patients [57], the horse should still be carefully monitored during and after the biopsy. However, 10 of 50 horses with clinicopathologic evidence of liver disease [13] showed clinical signs of epistaxis, bruising, and petechiae, although coagulation indices were not measured. In this same way, West [15] reported that 12 of 40 horses with liver disease evidenced mucosal petechiae or nosebleeds.

### 2.9. Triglycerides

In situations of negative energy balance, excessive movement of fatty acids out of fat tissue, leads to high blood TGs levels, although hepatic failure, secondary to lipidosis, rarely occurs unless there is visible lipemia in horses [58]; therefore, elevated TGs levels alone should not be used to diagnose hepatic lipidosis and liver failure.

### 2.10. Fibrinogen

Both in cases of parenchymal insufficiency and diffused intravascular coagulation (DIC) the concentrations of FIB can decrease. In addition, FIB is an acute phase protein indicative of acute inflammatory processes, so cholangiohepatitis can raise the levels of this analyte [59]. Indeed, low iron and high FIB plasma concentrations are both sensitive indicators of systemic inflammation in horses (sensitivity of 90 and 82%, respectively) [60]. Therefore, to correctly interpret blood FIB concentrations, it is important to consider the existence of concomitant liver failure [1]. Although it is rare even to develop hypofibrinogenemia in severe hepatopathies, dysfibrinogenemia, with the production of an abnormal fibrinogen molecule, is much more common [61] (Table 2).

## 3. Hematologic Abnormalities in Impaired Liver Function

Anemia or polycythemia may occur in horses with hepatic insufficiency. Anemia may be related to chronic inflammation in the presence of liver abscesses or neoplasia or with hemolysis. The anemia of chronic disease is caused by marrow suppression of red blood cell production. Inflammation also leads to a decrease in iron absorption, total iron binding capacity (TIBC)/transferrin, transferrin saturation, and an increase in ferritin to secure iron out of circulation. At the same time, inflammatory cytokines (predominantly IL-6) stimulate hepcidin production by the liver [62]. Hepcidin binds to the iron exporter (ferroportin) induced by the in-

**Table 3**

Common findings of hematological parameters associated with liver functional abnormalities [80–82].

Nonspecific Hematological Indicators of Liver Disease		
Hematocrit (%)	Anemia: Chronic inflammation Hemolysis Polycythemia: Secondary absolute erythrocytosis	32–48 (Hot-blooded) 24–44 (Cold-blooded)
White blood cells (10 <sup>3</sup> /μL)	Leukocytosis (infection or inflammation) Leukopenia (endotoxemia)	5.2–10.1
Platelets (10 <sup>3</sup> /μL)	Thrombocytopenia	100–300

hibition of iron entry into plasma from ferroportin-rich cells (enterocytes or macrophages). The resulting hypoferrremia is a common response to systemic infections or generalized inflammatory disorders that primarily serve as a defense mechanism by limiting the availability of iron to invading organisms [63]. Persistent hypoferrremia during inflammatory processes is the cause of anemia of chronic disease [64].

However, a rare fulminant intravascular hemolytic syndrome with prominent near-death hemoglobinuria has been reported in horses with hepatic failure in horses [65]. Although the mechanism by which liver pathology leads to intra-vascular hemolysis is unknown, BA values or its salts are considered hemolytic factors in horses [15]. In humans, bile salts bind to the outer layer of the lipid bilayer of the erythrocyte membrane, causing a preferential release of phospholipids and subsequent lysis [66].

In contrast, in horses that develop acute hepatic necrosis, hematocrit (PCV), iron concentrations, and percent iron saturation may remain elevated. Secondary absolute erythrocytosis results from increased stimulation of erythroid precursors as a physiological response to hypoxia from low ambient oxygen concentration, inadequate oxygen exchange due to pulmonary disease, abnormal hemoglobin affinity for oxygen, administration of exogenous substances like erythropoietin (EPO) or paraneoplastic production of EPO by tumors [67,68]. Two types of tumors associated with erythrocytosis in horses are hepatoblastoma (Beeler-Marfisi et al., 2000; [69–71]) and carcinoma ([72,73]; Beeler-Marfisi et al., 2000). Erythrocytosis in this type of neoplasm is usually associated with paraneoplastic production of EPO of tumor origin [74].

Leukocytosis with neutrophilia may be shifted to the right or the left. The right shift may accompany glucocorticoid-mediated stress [1]; the shift to the left responds to the systemic inflammatory response following the loss of Kupffer cells [1]. Examples of leukocytosis are cholelithiasis associated with recurrent colic [75], cholangiohepatitis [59], and other liver diseases [76]. In contrast, leukopenia with neutropenia is a common finding in infectious diseases directly related to the liver [10].

According to Durham et al. [1], erythrocytosis and leukocytosis are significant indicators of poor prognosis in horses with liver disease. The association between erythrocytosis and poor prognosis is probably related to dehydration in those cases with worse clinical status. Furthermore, these same investigators revealed an increased risk of non-survival in association with WBC > 10.0 × 10<sup>9</sup>/L compared to reference ranges.

Liver dysfunction can lead to thrombocytopenia from splenic sequestration in an enlarged spleen, secondary to portal hypertension or from decreased thrombopoietin production in the damaged liver [77]. In humans [78], and dogs [79], thrombocytopenia and impaired platelet function are hall-marks of chronic liver disease and cirrhosis. In horses these mechanisms are unknown. (Table 3)

#### 4. Conclusions

Biochemical and hematological analyses allow the diagnosis of different pathologies related to liver function in horses. The alteration of different biochemical analytes, like enzymes related to liver function, and the increase or decrease of different hematological parameters are not always specific to each type of dis-

ease. This is mainly because the excesses or defects of different parameters evaluated do not always occur in the same proportions. There are cases of altered liver function that can present a different magnitude of enzymatic or hematological modifications. Therefore, it is necessary, for a correct diagnosis, to carefully assess complete hematochemical variables, including the best-known enzyme activities, an expression and proof of correct and physiological liver function. In this way, veterinarians dedicated to the equine clinic can formulate a correct differential diagnosis of liver disorders from other diseases, and apply adequate treatment and prognosis.

#### Acknowledgments

The authors thank Dr Divers for his generosity in allowing us to use his tables for this review.

#### References

- [1] Durham AE, Newton JR, Smith KC, Hillyer MH, Hillyer LL, Smith MR, Marr CM. Retrospective analysis of historical, clinical, ultrasonographic, serum biochemical and haematological data in prognostic evaluation of equine liver disease. *Equine Vet J*. 2003;35:542–7.
- [2] Satué K, Gardon JC, Muñoz A. Use of laboratory testing to diagnose liver and biliary dysfunction in the horse. *J Gastroenterol Hepatol Res* 2013;2:807–13.
- [3] Theelen MJP, Beukers M, Grinwis GCM, Sloet van Oldruitenborgh-Oosterbaan MM. Chronic iron overload causing haemochromatosis and hepatopathy in 21 horses and one donkey. *Equine Vet J* 2019;51(3):304–9.
- [4] Tegtmeier B, Echelmeyer J, Pfankuche VM, Puff C, Todt D, Fischer N, Durham A, Feige K, Baumgärtner W, Steinmann E, Cavalleri JV. Chronic equine hepatitis infection in an adult gelding with severe hepatopathy. *Vet Med Sci* 2019;5(3):372–8.
- [5] Divers TJ, Tennant BC, Kumar A, McDonough S, Cullen J, Bhuvu N, Jain K, Chauhan LS, Scheel TKH, Lipkin WI, Laverack M, Trivedi S, Srinivasa S, Beard L, Rice CM, Burbelo PD, Renshaw RW, Dubovi E, Kapoor A. New parvovirus associated with serum hepatitis in horses after inoculation of common biological product. *Emerg Infect Dis* 2018;24(2):303–10.
- [6] Ramsauer AS, Badenhorst M, Cavalleri JV. Equine parvovirus hepatitis. *Equine Vet J* 2021;53(5):886–94.
- [7] Verhoef JNC, Allen AL, Harding JCS, Al-Dissi AN. Metallothionein expression in horses with chronic liver disease and its correlation with Ki-67 immunoreactivity. *Vet Pathol* 2018;55:703–10.
- [8] Divers TJ, Barton MH. Disorders of the Liver. In: Reed SM, Bayly WM, Sellon DC, editors. *Equine internal medicine*, 2018. St. Louis, Missouri: W.B. Saunders; 2018. p. 843–87.
- [9] Hughes KJ, McGorum BC, Love S, Dixon PM. Bilateral laryngeal paralysis associated with hepatic dysfunction and hepatic encephalopathy in six ponies and four horses. *Vet Rec* 2009;164:142–7.
- [10] Brazil T. 2012. Understanding and managing equine liver disease. Proceedings of the 18th Annual Meeting of the Italian Association of Equine Veterinarians (SIVE). Bologna (Italy), 2012; 8–10.
- [11] Hofmann AF. The continuing importance of bile acids in liver and intestinal disease. *Arch Intern Med* 1999;159:2647–58.
- [12] Dunkel B, Jones SA, Pinilla MJ. Serum bile acid concentrations, histopathological features, and short-, and long-term survival in horses with hepatic disease. *J Vet Intern Med* 2015;29(2):644–50.
- [13] McGorum BC, Murphy D, Love S. Clinicopathological features of equine primary hepatic disease: a review of 50 cases. *Vet Rec* 1999;145:134–9.
- [14] Delvescovo B, Tomlinson J, DeNotta S, Hodge E, Bookbinder L, Mohammed HO, Divers TJ. Bile acids, direct bilirubin and gamma-glutamyltransferase as prognostic indicators for horses with liver disease in the Eastern United States: 82 cases (1997–2019). *J Equine Vet Sci* 2021;105:103729.
- [15] West HJ. Clinical and pathological studies in horses with hepatic disease. *Equine Vet J* 1996;28:146–56.
- [16] Mendel VE, Witt MR, Gitchell BS, Gribble DN, Rogers QR, Segall HJ, Knight HD. Pyrrolizidine alkaloid-induced liver disease in horses: an early diagnosis. *Am J Vet Res* 1988;49:572–8.
- [17] Satué K, Miguel-Pastor L, Chicharro D, Gardón JC. Hepatic enzyme profile in horses. *Animals (Basel)* 2022;12:861.

- [18] Divers TJ. The equine liver in health and disease. *Proc Am Proc Am Assoc Equine Practit* 2015;6:66–103.
- [19] Mair TS, Divers TJ. Diseases of the liver and liver failure. In: Blikslager AT, White NA, Moore JN, Mair TS, editors. *The equine acute abdomen*, 2017. John Wiley & Sons, Inc.; 2017. p. 673–703.
- [20] Ankringa N, Wijnberg ID, Boerma S, Ijzer J. Copper-associated hepatic cirrhosis in a Friesian horse. *Tijdschr Diergeneesk* 2012;137:310–14.
- [21] Arfuso F, Giannetto G, Giudice E, Fazio F, Piccione G. Dynamic modulation of platelet aggregation, albumin and nonesterified fatty acids during physical exercise in Thoroughbred horses. *Res Vet Sci* 2016;104:86–91.
- [22] Schendl MJ, Redhead DN, Fearon KCH. The value of residual liver volume as a predictor of hepatic dysfunction and infection after major liver resection. *Gut* 2005;54:289–96.
- [23] Engelking LR. Evaluation of equine bilirubin and bile acid metabolism. *Compend Contin Educ Pract Vet* 1989;11:328–36.
- [24] Kaneko JJ, Rudolph WG, Wilson DV, Carlson GP. Bile acid fractionations by high performance liquid chromatography in equine liver disease. *Vet Res Commun* 1992;16:161–72.
- [25] Hoffmann WE, Baker G, Rieser S. Alterations in selected serum biochemical constituents in equids after induced hepatic disease. *Am J Vet Res* 1987;48:1343–7.
- [26] Divers TJ, Schappel KA, Sweeney RW. Persistent hyperbilirubinemia in a healthy thoroughbred horse. *Cornell Vet* 1993;83:237–42.
- [27] Barton MH. Disorders of the liver. In: Reed SM, Bayley WM, Sellon DC, editors. *Equine internal medicine*, 2010. St. Louis: Saunders; 2010. p. 939–75.
- [28] DeNotta SL, Divers TJ. Clinical pathology in the adult sick horse: the gastrointestinal system and liver. *Vet Clin North Am Equine Pract* 2020;36:105–20.
- [29] Dunkel B, Chaney KP, Dallap-Schaer BL. Putative intestinal hyperammonemia in horses: 36 cases. *Equine Vet J* 2011;43:133–40.
- [30] Mair TS, Love S. *Gastroenterology 2. Hepatic and intestinal disorders*. Equine Med, Surg Reprod 2012:49–65.
- [31] Divers TJ. Liver failure, anemia, and blood transfusion. In: Orsini JA, Divers TJ, editors. *Equine emergencies treatments and procedures*, 2014. St. Louis, Missouri: Elsevier; 2014. p. 268–88.
- [32] Mair TS, Jones RD. Acute encephalopathy and hyperammonaemia in a horse without evidence of liver disease. *Vet Rec* 1995;137:642–3.
- [33] Peek SF, Divers TJ, Jackson CJ. Hyperammonemia associated with encephalopathy and abdominal pain without evidence of liver disease in four mature horses. *Equine Vet J* 1997;29:70–4.
- [34] Sharkey LC, DeWitt S, Stockman C. Neurologic signs and hyperammonemia in a horse with colic. *Vet Clin Pathol* 2006;35:254–8.
- [35] McCornico RS, Duckett WM, Wood PA. Persistent hyperammonemia in two related Morgan weanlings. *J Vet Intern Med* 1997;11:264–6.
- [36] Campollo O, Sprengers D, McIntyre N. The BCAA/AAA ratio of plasma amino acids in three different groups of cirrhotics. *Rev Invest Clin* 1992;44:513–518.
- [37] Schnabel LV, Njaa BL, Gold JR, Meseck EK. Primary alimentary lymphoma with metastasis to the liver causing encephalopathy in a horse. *J Vet Intern Med* 2006;20:204–6.
- [38] Oliveira-Filho JP, Cagnin DQ, Badial PR, Pessoa MA, Del Piero F, Borges AS. Hepatoencephalopathy syndrome due to *Cassia occidentalis* (leguminosae Caesalpinioideae) seed ingestion in horses. *Equine Vet J* 2013;45:240–4.
- [39] Tajiri K, Shimizu Y. Branched-chain amino acids in liver diseases. *World J Gastroenterol* 2013;19(43):7620–9.
- [40] Tennant BC. Hepatic function. In: Kaneko JJ, Harvey JW, Bruss ML, editors. *Clinical biochemistry of domestic animals*, 1997. Maryland Heights (MO): Elsevier Academic Press; 1997. p. 379–413.
- [41] Francoz C, Prié D, Abdelrazek W, Moreau R, Mandot A, Belghiti J, Valla D, Durand F. Inaccuracies of creatinine and creatinine-based equations in candidates for liver transplantation with low creatinine: impact on the model for end-stage liver disease score. *Liver Transpl* 2010;16:1169–77.
- [42] Savage VL, Marr CM, Bailey M, Smith S. Prevalence of acute kidney injury in a population of hospitalized horses. *J Vet Intern Med* 2019;33(5):2294–2301.
- [43] Axon JE, Palmer JE. Clinical pathology of the foal. *Vet Clin North Am: Equine Pract*. 2008;24:357–85.
- [44] Bauer JE, Asquith RL, Kivipelto J. Serum biochemical indicators of liver function in neonatal foals. *Am J Vet Res* 1989;50:2037–41.
- [45] Sellon DC, Spoulding K, Breuhaus BA, Katz L, Mealey R. Hepatic abscesses in three horses. *J Am Vet Med Assoc* 2000;216:882–7.
- [46] Parraga ME, Carlson GP, Thurmond M. Serum protein concentrations in horses with severe liver disease: a retrospective study and review of the literature. *J Vet Intern Med* 1995;9:154–61.
- [47] Badial PR, Tallmadge RL, Miller S, Stokol T, Richards K, Borges AS, Felipe MJ. Applied protein and molecular techniques for characterization of B cell neoplasms in horses. *Clin Vaccine Immunol* 2015;22(11):1133–45.
- [48] Last RD, Hill JM, Vorster JH, Bosch SJ, Griffiths C. *Vetdiagnostix laboratory manual*. 2nd ed. Pretoria: Serrati Publishers; 2010. AC/0492-10.
- [49] Arfuso F, Giannetto C, Fazio F, Panzera F, Piccione G. Training program intensity induces an acute phase response in clinically healthy horses. *J Equine Vet Sci* 2020;88:102986.
- [50] Arfuso F, Giudice E, Di Pietro S, Piccione G, Giannetto C. Modulation of serum protein electrophoretic pattern and leukocyte population in horses vaccinated against West Nile virus. *Animals* 2021;11(2):477.
- [51] Piccione G, Arfuso F, Marafioti S, Giannetto C, Giudice E, Fazio F. Different training schedules influence serum electrophoretic protein profile in the athletic horse. *J Equine Vet Sci* 2015;35:856–9.
- [52] Johnstone LK, Engiles JB, Aceto H, Buechner-Maxwell V, Divers T, Gardner R, Levine R, Scherrer N, Tewari D, Tomlinson J, Johnson AL. Retrospective evaluation of horses diagnosed with neuroborreliosis on postmortem examination: 16 cases (2004–2015). *J Vet Intern Med* 2016;30(4):1305–12.
- [53] Pecoraro HL, Felipe MJB, Miller AD, Divers TJ, Simpson KW, Guyer KM, Duhamel GE. Neuroborreliosis in a horse with common variable immunodeficiency. *J Vet Diagn Invest* 2019;31(2):241–5.
- [54] Aleman M, Costa LRR, Crowe C. Presumed neuroglycopenia caused by severe hypoglycemia in horses. *Vet Intern Med* 2018;32(5):1731–9.
- [55] Tomlinson JE, Kapoor A, Kumar A. Viral testing of 18 consecutive cases of equine serum hepatitis: a prospective study (2014–2018). *J Vet Intern Med* 2019;33(1):251–7.
- [56] Peek SF, Divers TJ. Medical treatment of cholangiohepatitis and cholelithiasis in mature horses: 9 cases (1991–1998). *Equine Vet J* 2000;32:301–306.
- [57] Johns IC, Sweeney RW. Coagulation abnormalities and complications after percutaneous liver biopsy in horses. *J Vet Intern Med* 2008;22:185–9.
- [58] Dunkel B, McKenzie HC. Severe hypertriglyceridaemia in clinically ill horses: diagnosis, treatment and outcome. *Equine Vet J* 2003;35:590–5.
- [59] Peek SF. Cholangiohepatitis in the mature horse. *Equine Vet Educ* 2004;16:72–5.
- [60] Corradini I, Armengou L, Viu J, Rodríguez-Pozo ML, Cesarini C, José-Cuñilleras E. Parallel testing of plasma iron and fibrinogen concentrations to detect systemic inflammation in hospitalized horses. *J Vet Emerg Crit Care* 2014;24:414–20.
- [61] Kujovich J. Hemostatic defects in end-stage liver disease. *Crit Care Clin* 2005;21:563–87.
- [62] Ganz T. The discovery of the iron-regulatory hormone Hephcidin. *Clin Chem* 2019;65:1330–1.
- [63] Ganz T, Nemeth E. Iron imports. IV. Hephcidin and regulation of body iron metabolism. *Am J Physiol Gastrointest Liver Physiol* 2006;290:G199–G203.
- [64] Nemeth E, Ganz T. Anemia of inflammation. *Hematol Oncol Clin North Am* 2014;28:671–81.
- [65] Ramaiah SK, Harvey JW, Giguere S, Franklin RP, Crawford PC. Intravascular hemolysis associated with liver disease in a horse with marked neutrophil hypersegmentation. *J Vet Intern Med* 2003;17:360–3.
- [66] Martin GP, el Hariri LM, Marriott C. Bile salt and lysophosphatidylcholine-induced membrane damage in human erythrocytes. *J Pharm Pharmacol* 1992;44:646–50.
- [67] Koch TG, Wen X, Bienzle D. Lymphoma, erythrocytosis, and tumor erythropoietin gene expression in a horse. *J Vet Intern Med* 2006;20:1251–5.
- [68] McMullin MF, Bareford D, Campbell P, Green AR, Harrison C, Hunt B, Oscier D, Polkey MI, Reilly JT, Rosenthal E, Ryan K, Pearson TC, Wilkins B. General haematology task force of the british committee for standards in haematology. guidelines for the diagnosis, investigation and management of polycythaemia/erythrocytosis. *Br J Haematol* 2005;130:174–95.
- [69] Axon JE, Russell CM, Begg AP, Adkins AR. Erythrocytosis and pleural effusion associated with a hepatoblastoma in a Thoroughbred yearling. *Aust Vet J* 2008;86:329–33.
- [70] Lennox TJ, Wilson JH, Hayden DW, Bouljihad M, Sage AM, Walsler MM, Manivel JC. Hepatoblastoma with erythrocytosis in a young female horse. *J Am Vet Med Assoc* 2000;216:718–21.
- [71] Tirosh-Levy S, Perl S, Valentine BA, Kelmer G. Erythrocytosis and fatigue fractures associated with hepatoblastoma in a 3-year-old gelding. *J S Afr Vet Assoc* 2019;28:90:e1–e5.
- [72] Roby KA, Beech J, Bloom JC, Black M. Hepatocellular carcinoma associated with erythrocytosis and hypoglycaemia in a yearling filly. *J Am Vet Med Assoc* 1990;196:465–7.
- [73] Cook G, Divers TJ, Rowland PH. Hypercalcaemia and erythrocytosis in a mare associated with a metastatic carcinoma. *Equine Vet J* 1995;27:316–18.
- [74] McFadzean AJ, Todd D, Tso SC. Erythrocytosis associated with hepatocellular carcinoma. *Blood* 1967;29:808–11.
- [75] Ryu SH, Bak UB, Lee CW, Lee YL. Cholelithiasis associated with recurrent colic in a Thoroughbred mare. *J Vet Sci* 2004;5:79–82.
- [76] Borchers A, Magdesian KG, Holland S, Pusterla N, Wilson WD. Successful treatment and polymerase chain reaction (PCR) confirmation of Tyzzer's disease in a foal and clinical and pathologic characteristics of 6 additional foals (1986–2005). *J Vet Intern Med* 2006;20:1212–18.
- [77] Lisman T, Porte RJ. Rebalanced hemostasis in patients with liver disease: Evidence and clinical consequences. *Blood* 2010;116:878–85.
- [78] Witters P, Freson K, Verslype C, Peerlinck K, Hoylaerts M, Nevens F, Van Geet C, Cassiman D. Review article: blood platelet number and function in chronic liver disease and cirrhosis. *Aliment Pharmacol Ther* 2008;27:1017–29.
- [79] Lester C, Cooper J, Peters RM, Webster CR. Retrospective evaluation of acute liver failure in dogs (1995–2012): 49 cases. *J Vet Emerg Crit Care (San Antonio)* 2016;26:559–67.
- [80] Satué K, Muñoz A, Gardón JC. Interpretation of alterations in the horse erythrogram. *J Hematol Res* 2014;1:1–10.
- [81] Satué K, Muñoz A, Gardón JC. Interpretation of the equine leukogram. *J Hematol Res* 2014;1:27–35.
- [82] Satué K, Gardón JC, Muñoz A. Interpretation of platelets in the horse. *J Hematol Res* 2017;4:19–25.
- [83] Barton MH, LeRoy BE. Serum bile acids concentrations in healthy and clinically ill neonatal foals. *J Vet Intern Med* 2007;21:508–13.